

Algorithms for de novo genome assembly and disease analytics

Michael Schatz

March 11, 2014

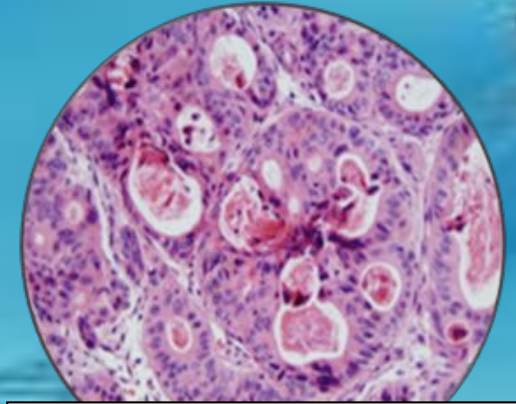
University of Florida Genetics Institute



Schatz Lab Overview



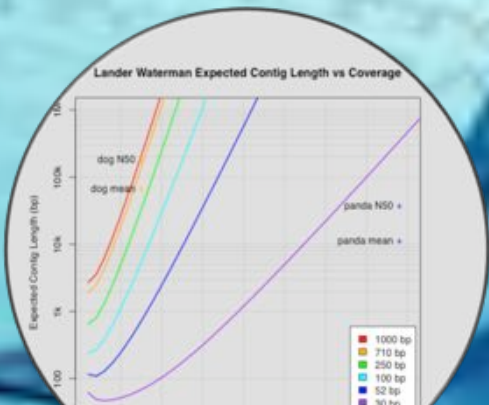
Computation



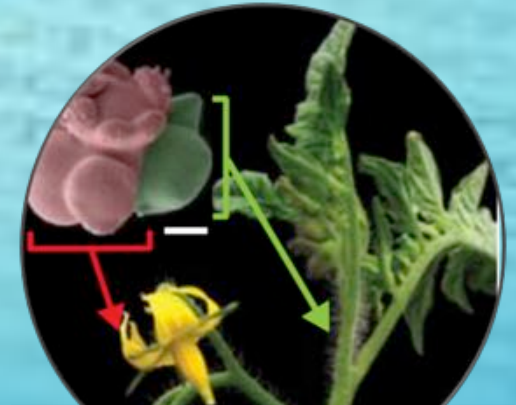
Human Genetics



Sequencing



Modeling



Plant Genomics

Introductions



Ke Jiang

Transcriptomics and
epigenetics

Tomato &
Solanaceae



**Srividya "Sri"
Ramakrishnan**

DOE Systems Biology
Knowledgebase

Worlds fastest -omics
pipelines



Maria Nattestad

Hi-C Chromatin
Interactions

Plant Assembly &
Analysis



Tyler Garvin

CNV analysis of
single cells

Breast & Prostate
Cancer

Outline

1. De novo assembly by analogy
2. Long Read Assembly
3. Disease Analytics



Shredded Book Reconstruction

- Dickens accidentally shreds the first printing of A Tale of Two Cities
 - Text printed on 5 long spools

It was	the	best	of	times,	it	was	the	worst	of	times,	it	was	the	age	of	wisdom,	it	was	the	age	of	foolishness, ...	
It was	the	best	of	times,	it	was	the	worst	of	times,	it	was	the	age	of	wisdom,	it	was	the	age	of	foolishness, ...	
It was	the	best	of	times,	it	was	the	worst	of	times,	it	was	the	age	of	wisdom,	it	was	the	age	of	foolishness, ...	
It was	the	best	of	times,	it	was	the	worst	of	times,	it	was	the	age	of	wisdom,	it	was	the	age	of	foolishness, ...	
It	was	the	best	of	times,	it	was	the	worst	of	times,	it	was	the	age	of	wisdom,	it	was	the	age	of	foolishness, ...

- How can he reconstruct the text?
 - 5 copies x 138,656 words / 5 words per fragment = 138k fragments
 - The short fragments from every copy are mixed together
 - Some fragments are identical

Greedy Reconstruction

It was the best of
age of wisdom, it was
best of times, it was
it was the age of
it was the age of
it was the worst of
of times, it was the
of times, it was the
of wisdom, it was the
the age of wisdom, it
the best of times, it
the worst of times, it
times, it was the age
times, it was the worst
was the age of wisdom,
was the age of foolishness,
was the best of times,
was the worst of times,
wisdom, it was the age
worst of times, it was

It was the best of
was the best of times,
the best of times, it
best of times, it was
of times, it was the
of times, it was the
times, it was the worst
times, it was the age

The repeated sequence make the correct reconstruction ambiguous

- It was the best of times, it was the [worst/age]

Model the assembly problem as a graph problem

de Bruijn Graph Construction

- $D_k = (V, E)$
 - $V =$ All length- k subfragments ($k < l$)
 - $E =$ Directed edges between consecutive subfragments
 - Nodes overlap by $k-1$ words

Original Fragment

It was the best of

Directed Edge

It was the best → was the best of

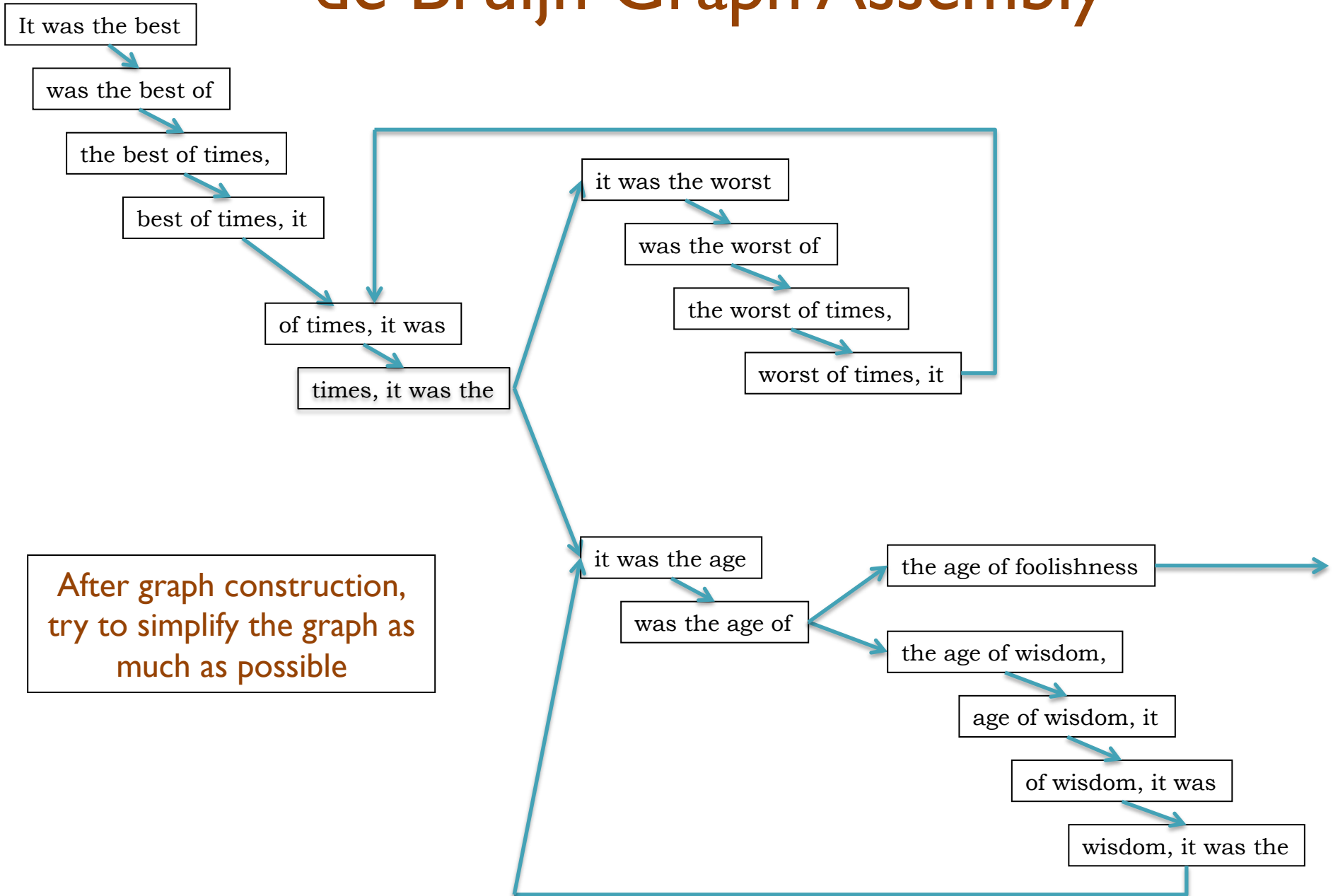
- Locally constructed graph reveals the global sequence structure
 - Overlaps between sequences implicitly computed

de Bruijn, 1946

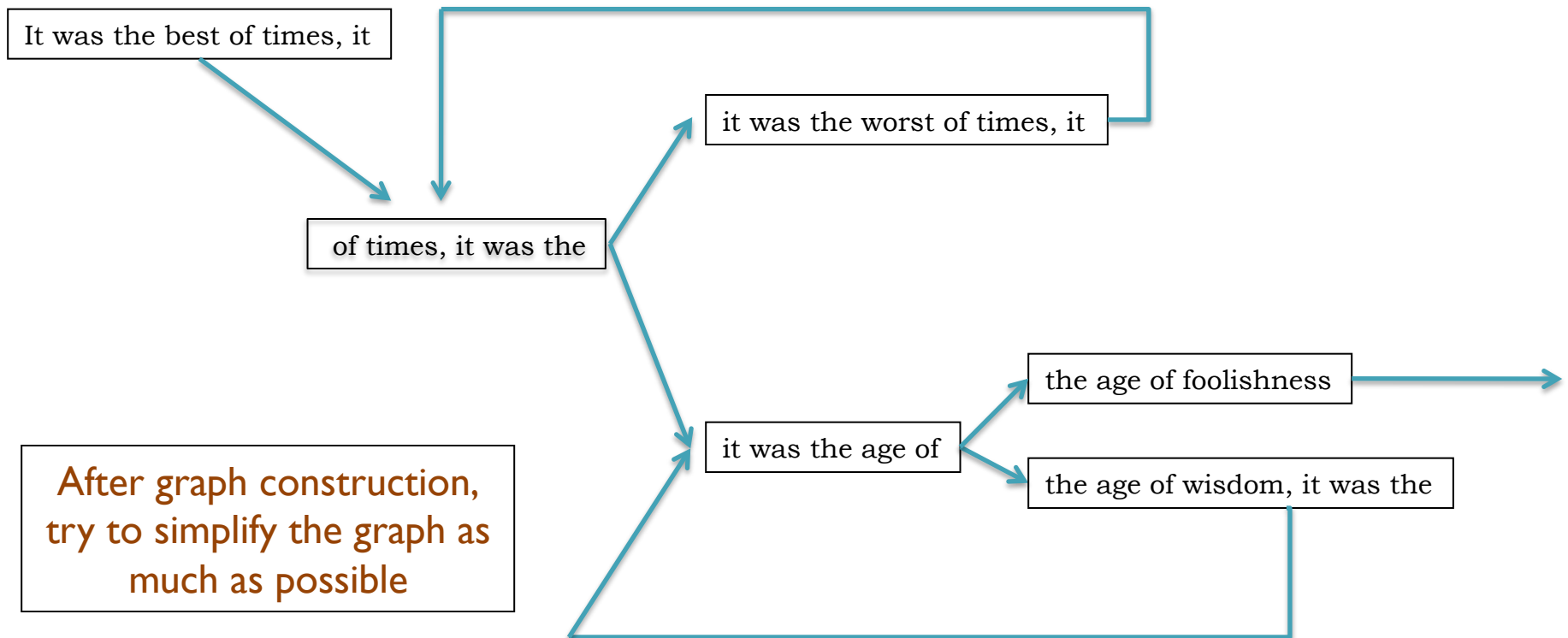
Idury and Waterman, 1995

Pevzner, Tang, Waterman, 2001

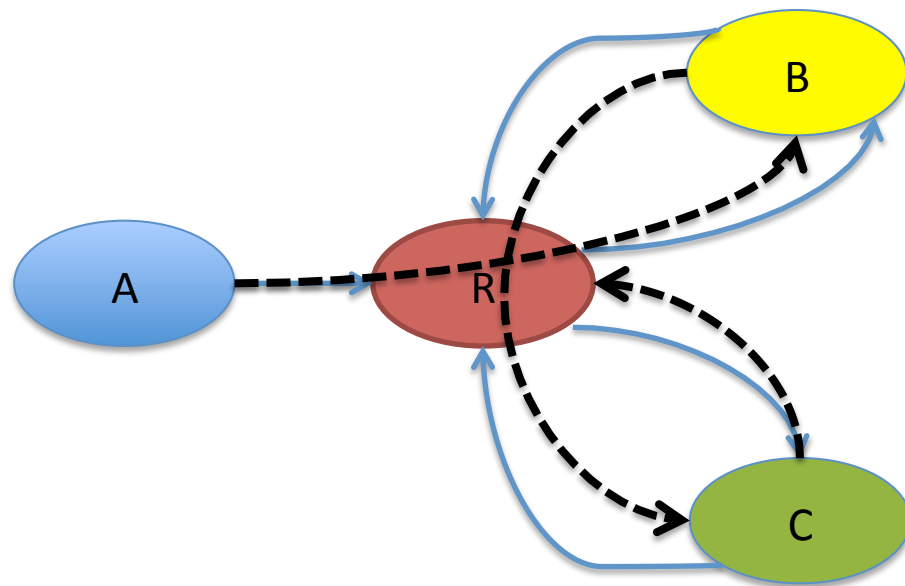
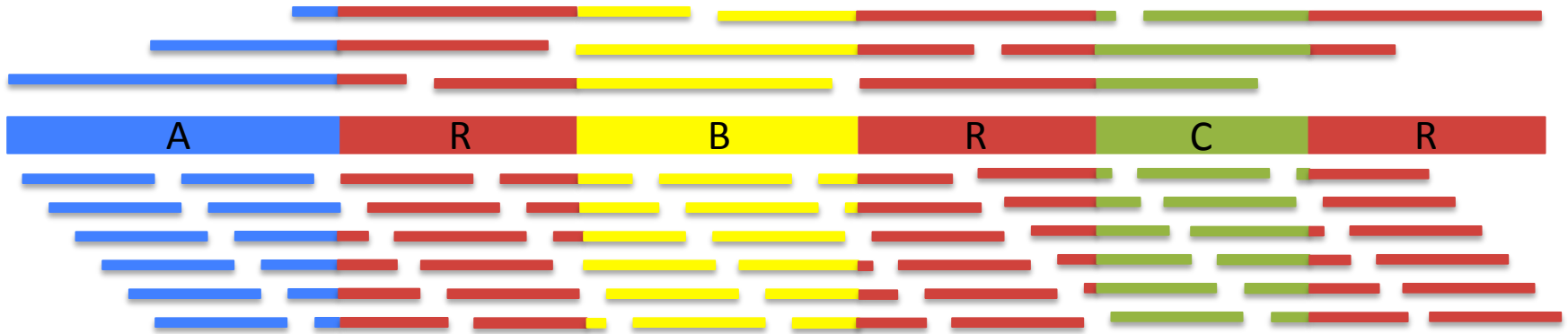
de Bruijn Graph Assembly



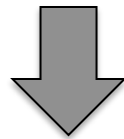
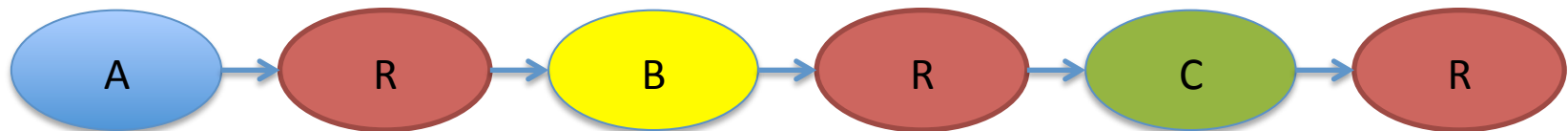
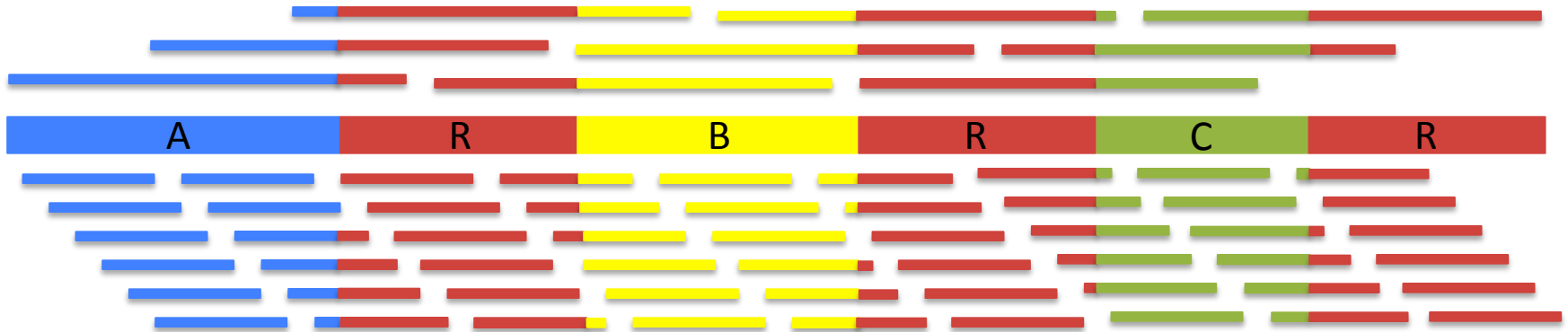
de Bruijn Graph Assembly



Assembly Complexity



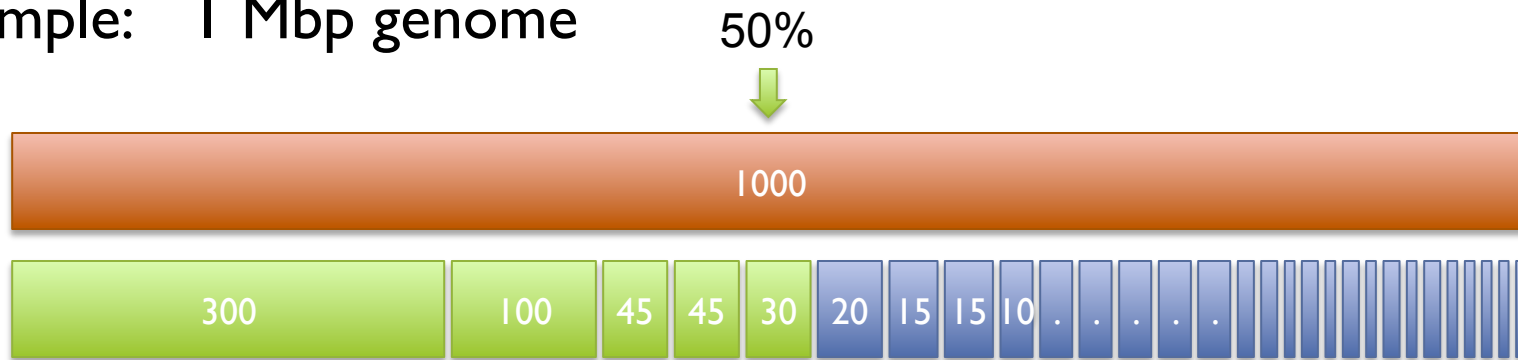
Assembly Complexity



N50 size

Def: 50% of the genome is in contigs as large as the N50 value

Example: 1 Mbp genome



N50 size = 30 kbp

(300k+100k+45k+45k+30k = 520k \geq 500kbp)

Note:

A “good” N50 size is a moving target relative to other recent publications. 10-20kbp contig N50 is currently a typical value for most “simple” genomes.

Outline

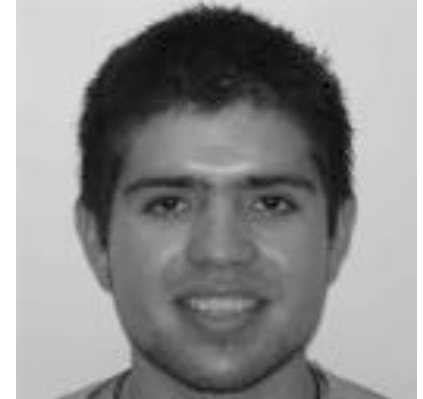
1. De novo assembly by analogy
2. Long read assembly
3. Disease Analytics



Population structure of *Oryza sativa*

Conclusions

- Very high quality representation of the “gene-space”
 - Overall identity ~99.9%
 - Less than 1% of exonic bases missing
- Genome-specific genes enriched for disease resistance
 - Reflects their geographic and environmental diversity
 - Detailed analysis of agriculturally important loci
- Assemblies fragmented at (high copy) repeats
 - Missing regions have mean kmer coverage >10,000x
 - Difficult to identify full length gene models and regulatory features

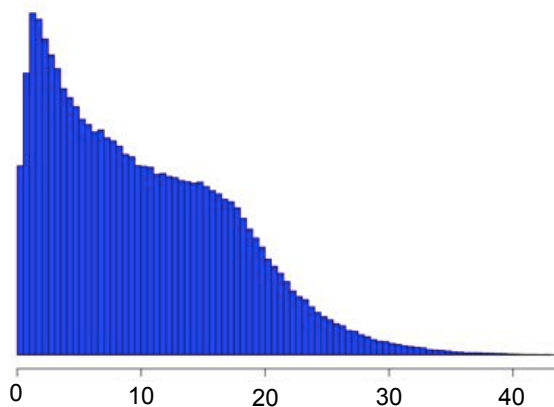


New whole genome de novo assemblies of three divergent strains of rice (*O. sativa*) documents novel gene space of *aus* and *indica*

Schatz, MC, Maron, L, Stein, J, Ware, DW, McCouch, S, McCombie WR et al (2014) *In preparation*

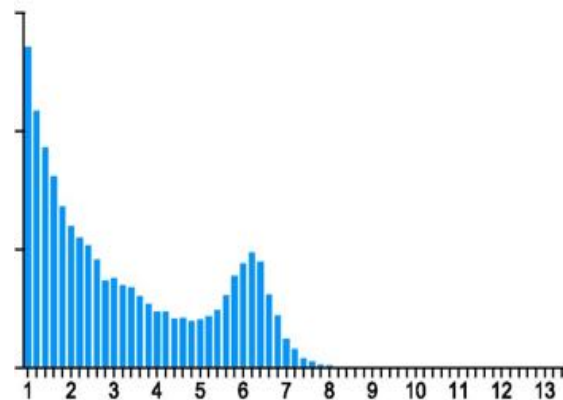
Long Read Sequencing Technology

PacBio RS II



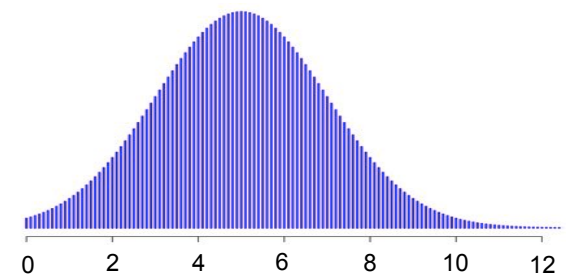
CSHL/PacBio

Moleculo



(Voskoboynik et al. 2013)

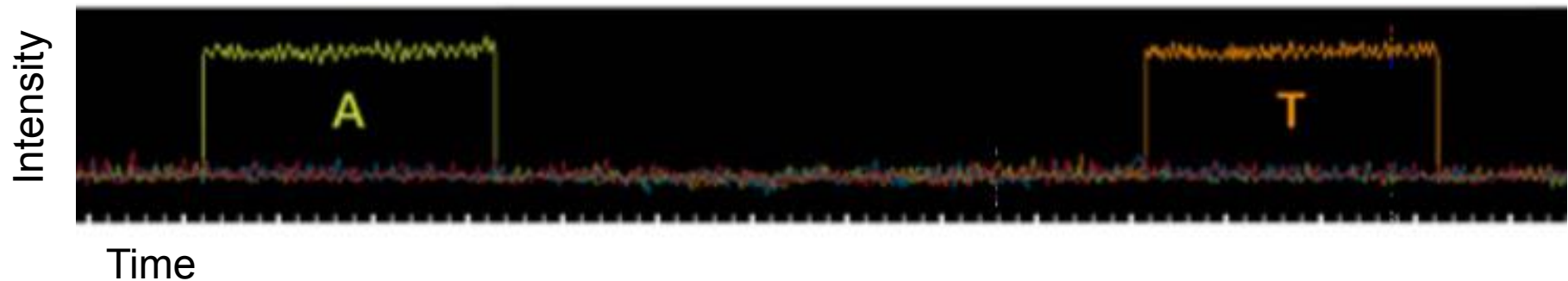
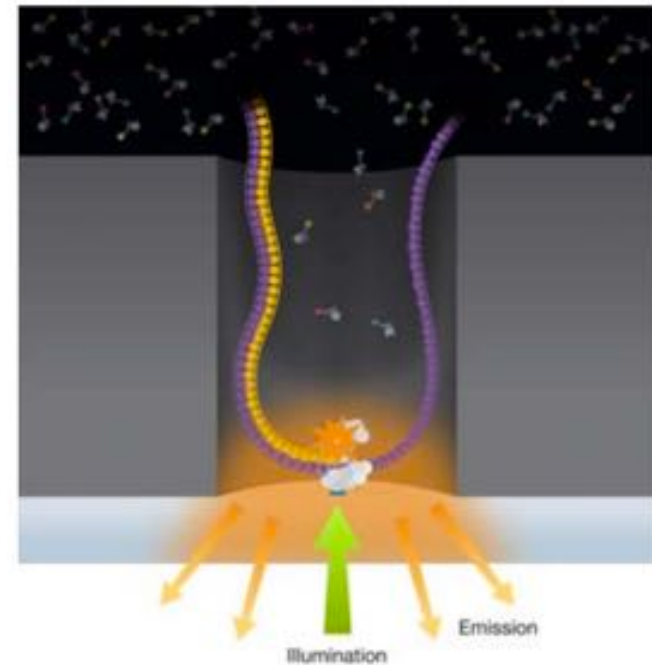
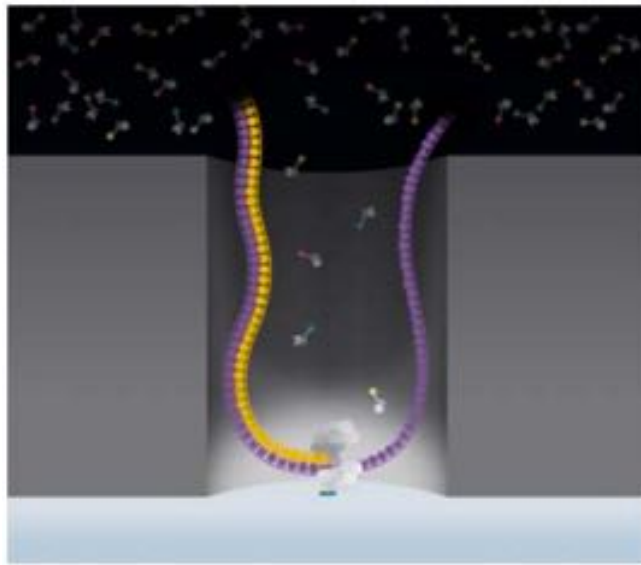
Oxford Nanopore



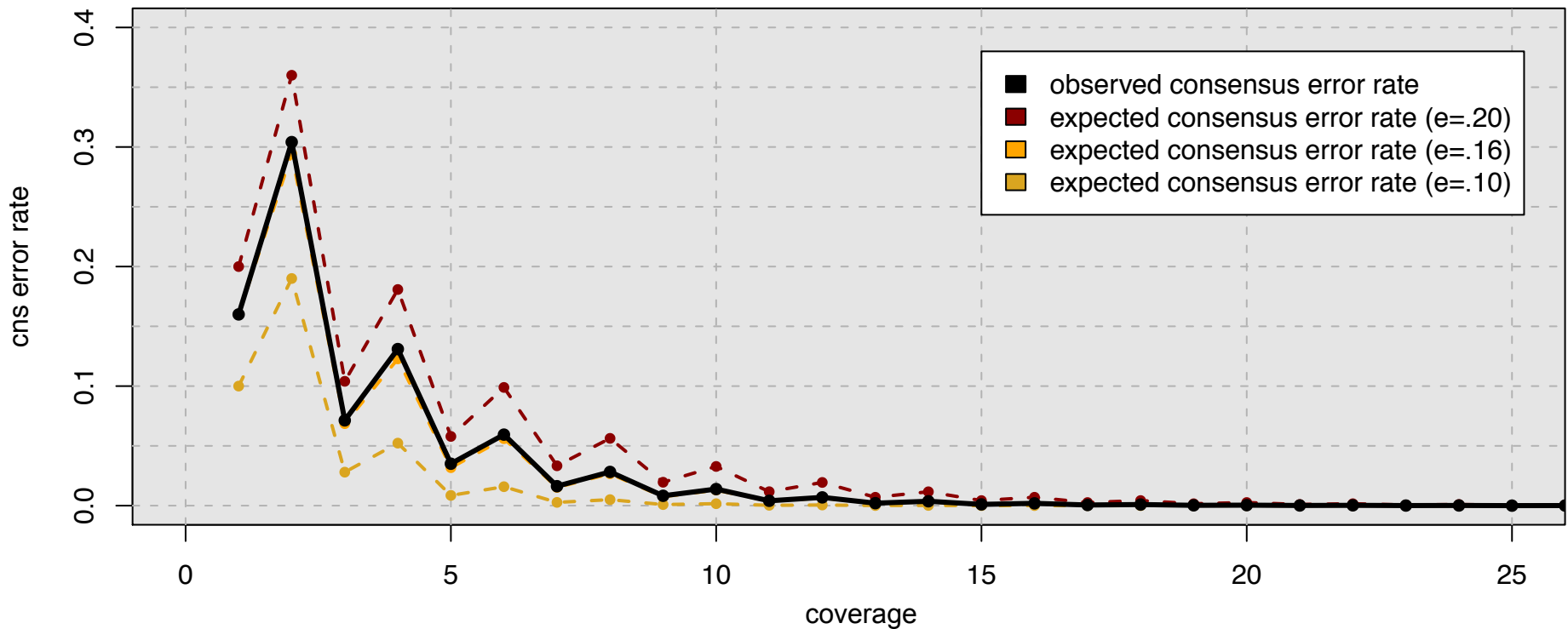
Broad/OxNano @ AGBT ***

SMRT Sequencing

Imaging of fluorescently phospholinked labeled nucleotides as they are incorporated by a polymerase anchored to a Zero-Mode Waveguide (ZMW).



Consensus Accuracy and Coverage



Coverage can overcome random errors

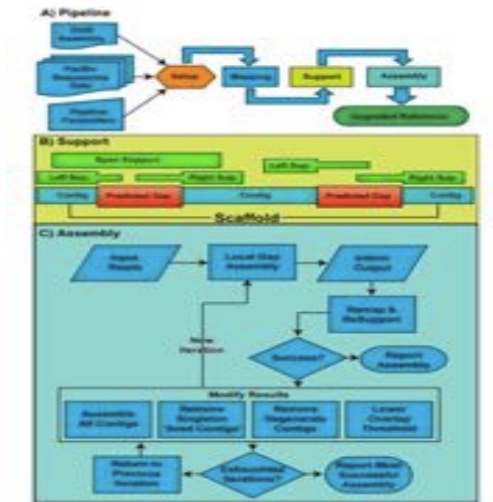
- Dashed: error model from binomial sampling
- Solid: observed accuracy

Koren, Schatz, et al (2012)
Nature Biotechnology. 30:693–700

$$CNS\ Error = \sum_{i=\lfloor c/2 \rfloor}^c \binom{c}{i} (e)^i (1-e)^{n-i}$$

PacBio Assembly Algorithms

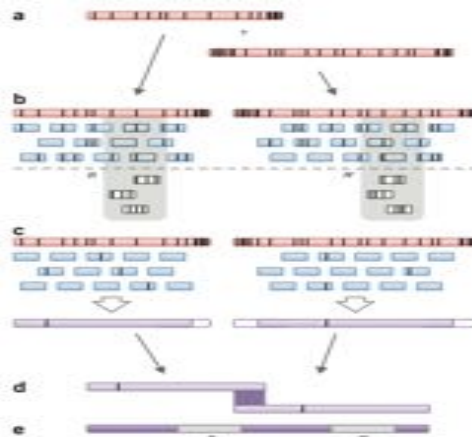
PBJelly



**Gap Filling
and Assembly Upgrade**

English *et al* (2012)
PLOS One. 7(11): e47768

PacBioToCA & ECTools



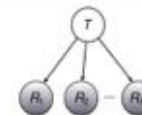
**Hybrid/PB-only Error
Correction**

Koren, Schatz, *et al* (2012)
Nature Biotechnology. 30:693–700

HGAP & Quiver



$$\Pr(\mathbf{R} | T) = \prod_k \Pr(R_k | T)$$



Quiver Performance Results Comparison to Reference Genome (<i>M. ruber</i> ; 3.1 MB; SMRT® Cells)		
	Initial Assembly	Quiver Consensus
QV	43.4	54.5
Accuracy	99.99540%	99.99964%
Differences	141	11

**PB-only Correction &
Polishing**

Chin *et al* (2013)
Nature Methods. 10:563–569

< 5x

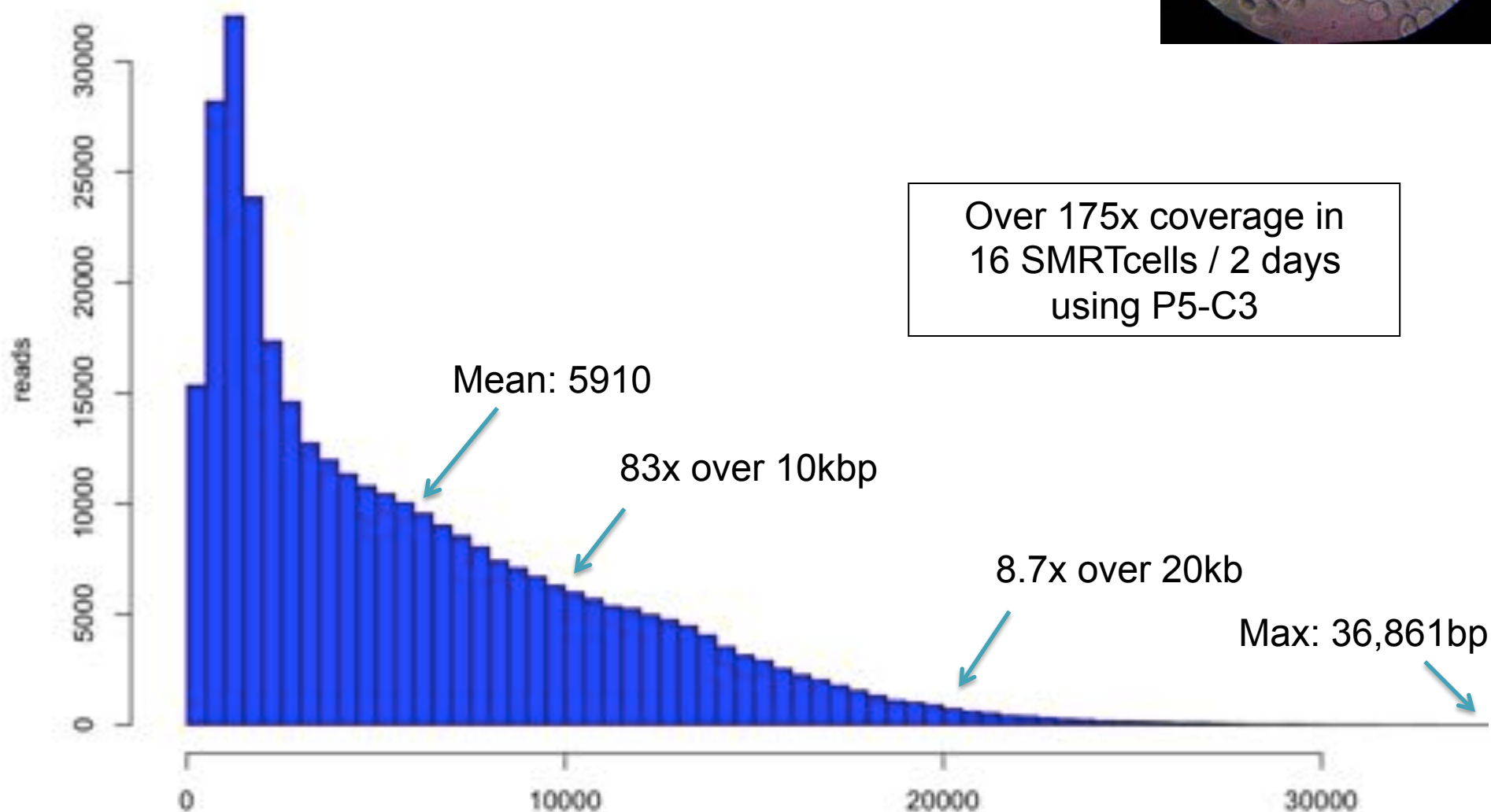
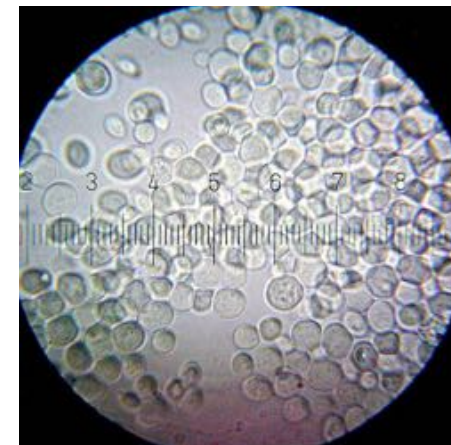
PacBio Coverage

> 50x

S. cerevisiae W303

PacBio RS II sequencing at CSHL by Dick McCombie

- Size selection using an 7 Kb elution window on a BluePippin™ device from Sage Science



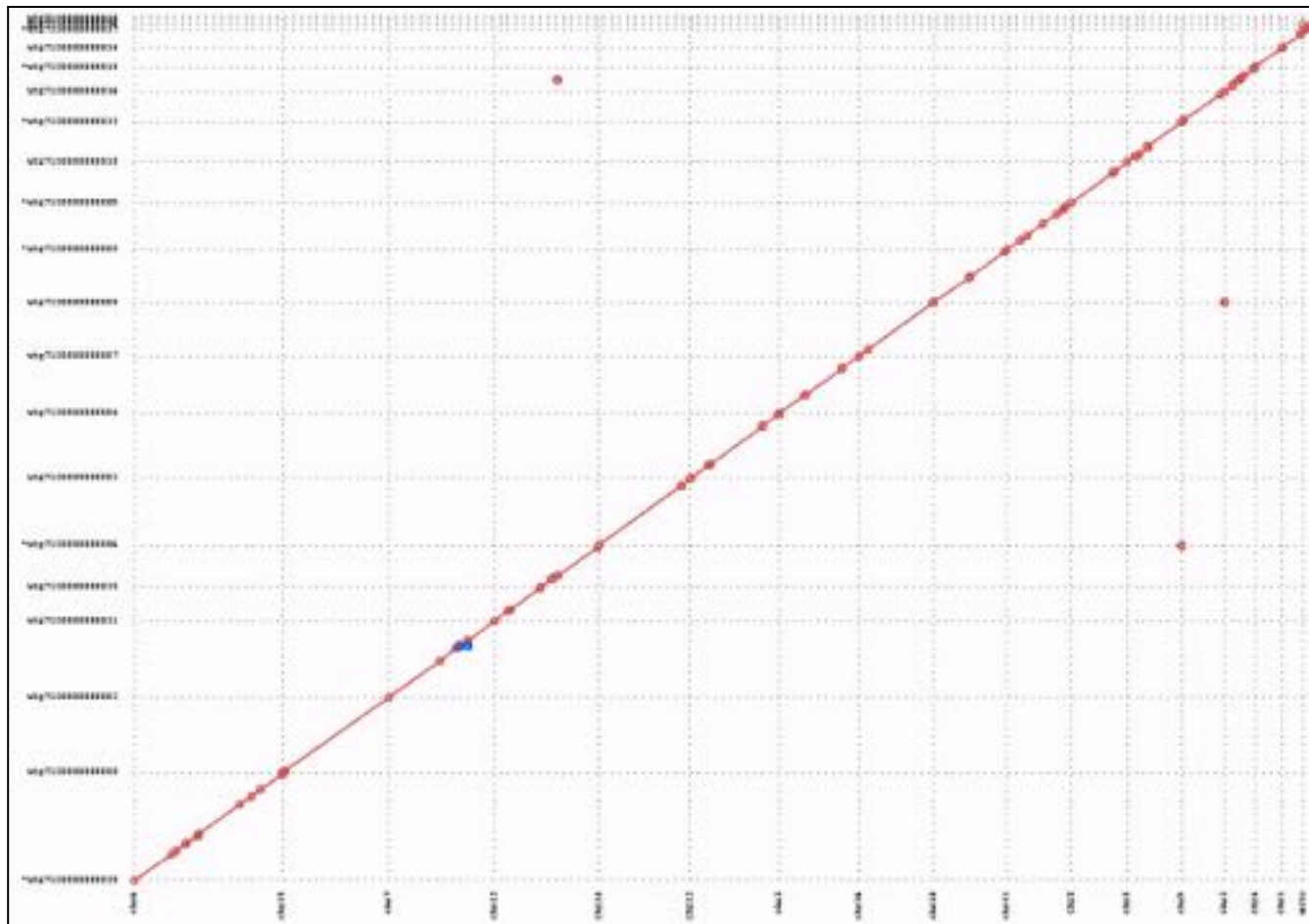
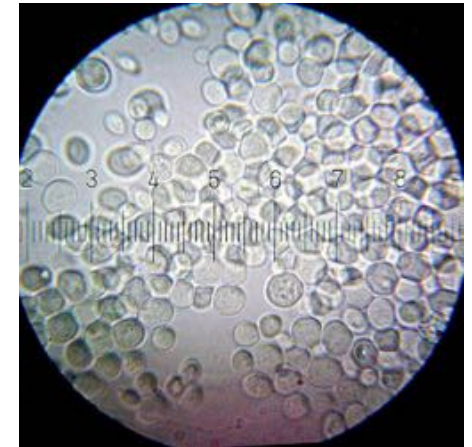
S. cerevisiae W303

S288C Reference sequence

- 12.1Mbp; 16 chromo + mitochondria; N50: 924kbp

PacBio assembly using HGAP + Celera Assembler

- 12.4Mbp; 21 non-redundant contigs; N50: 811kbp; >99.8% id



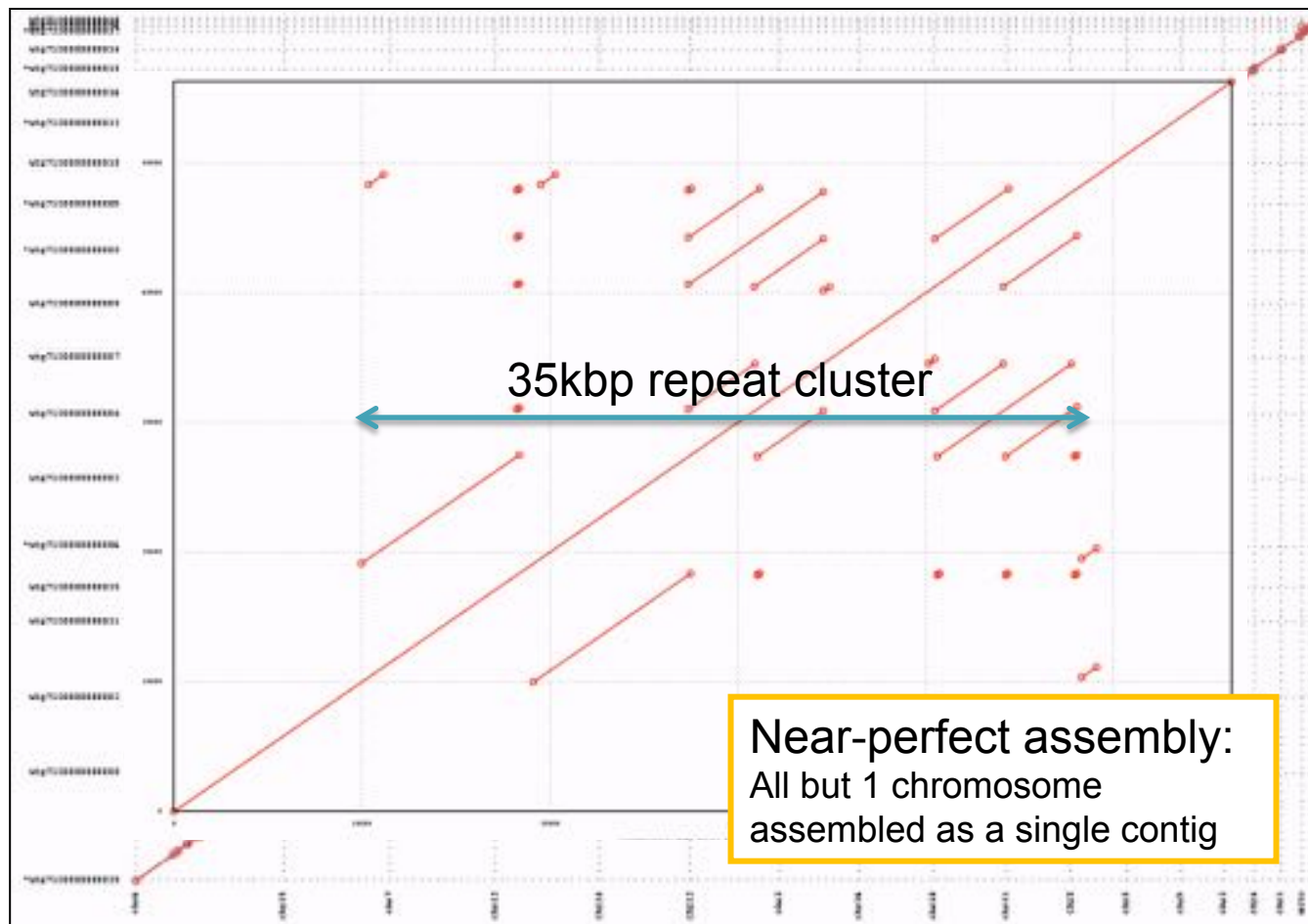
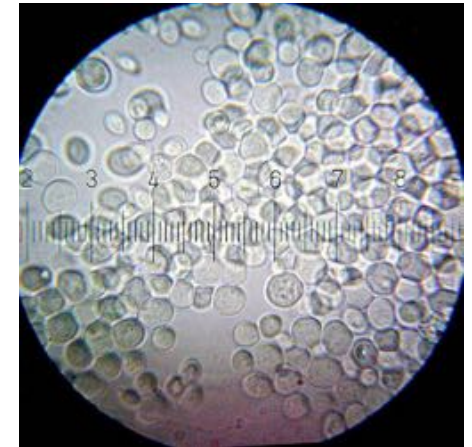
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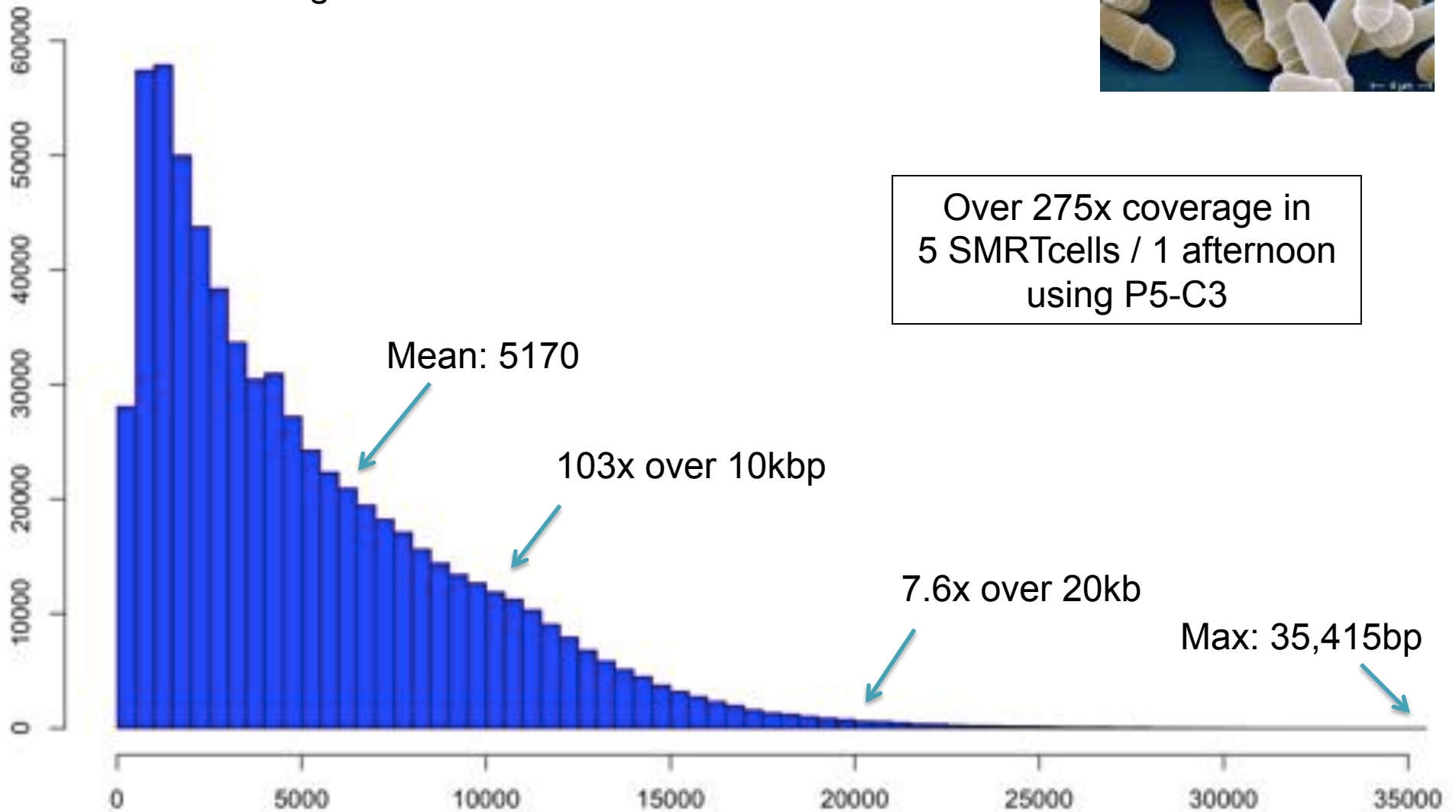
- 12.4Mbp; 21 non-redundant contigs; N50: 811kbp; >99.8% id



S. pombe dg2 I

PacBio RS II sequencing at CSHL

- Size selection using a 7 Kb elution window on a BluePippin™ device from Sage Science



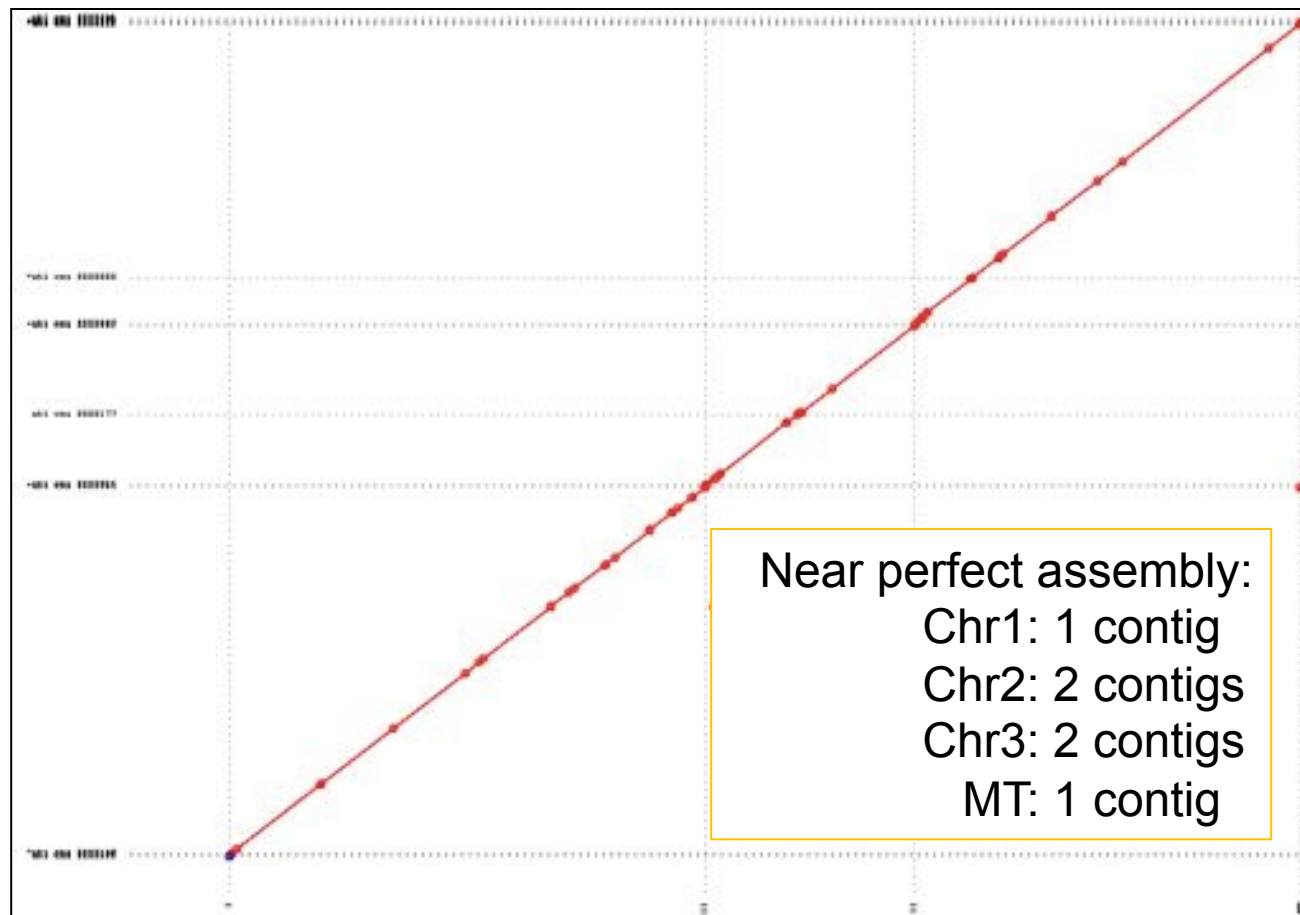
S. pombe dg2 I

ASM294 Reference sequence

- 12.6Mbp; 3 chromo + mitochondria; N50: 4.53Mbp

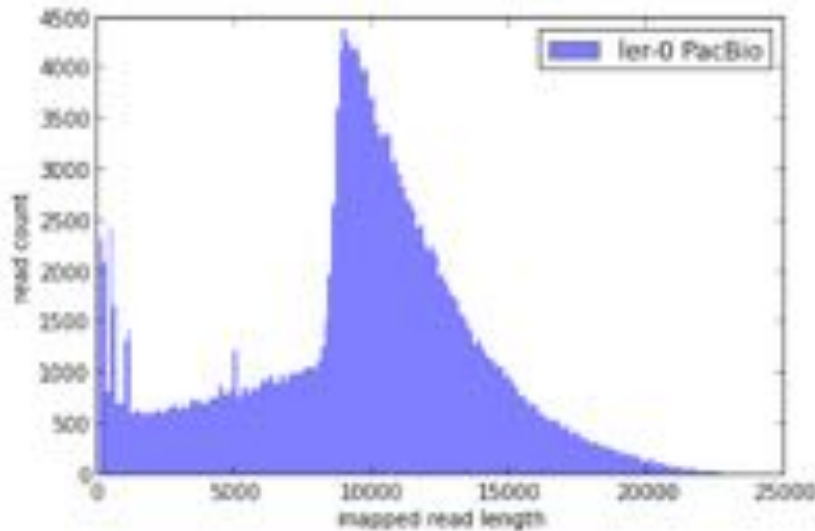
PacBio assembly using HGAP + Celera Assembler

- 12.7Mbp; 13 non-redundant contigs; N50: 3.83Mbp; >99.9% id



A. thaliana Ler-0

<http://blog.pacificbiosciences.com/2013/08/new-data-release-arabidopsis-assembly.html>



A. thaliana Ler-0 sequenced at PacBio

- Sequenced using the previous P4 enzyme and C2 chemistry
- Size selection using an 8 Kb to 50 Kb elution window on a BluePippin™ device from Sage Science
- Total coverage >119x

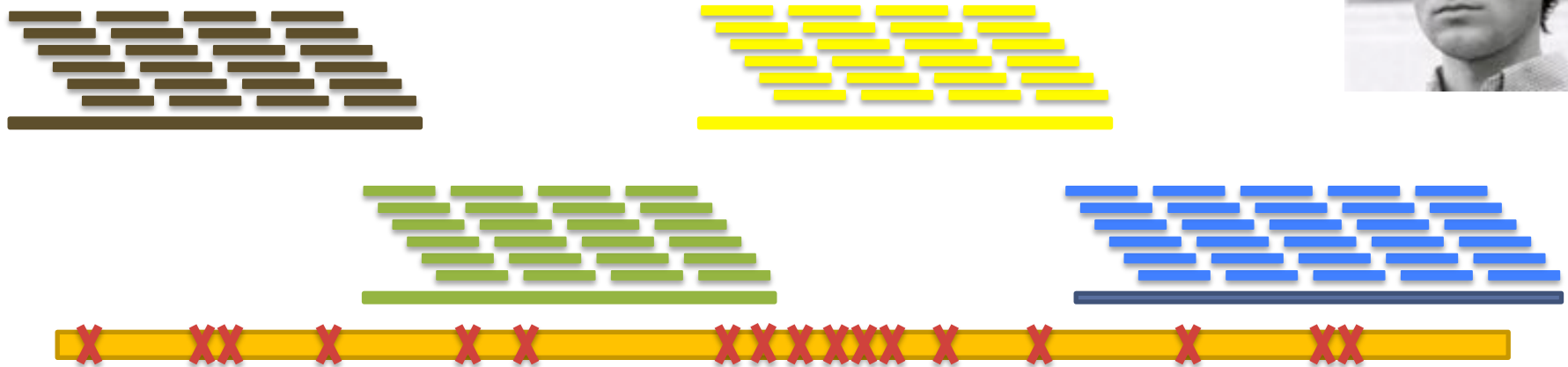
Genome size: 124.6 Mbp
Chromosome N50: 23.0 Mbp
Corrected coverage: 20x over 10kb

Sum of Contig Lengths: 149.5Mb
N50 Contig Length: 8.4 Mb
Number of Contigs: 1788

High quality assembly of chromosome arms
Assembly Performance: $8.4\text{Mbp}/23\text{Mbp} = 36\%$
MiSeq assembly: $63\text{kbp}/23\text{Mbp} = .2\%$

ECTools: Error Correction with pre-assembled reads

<https://github.com/jgurtowski/ectools>



Short Reads -> Assemble Unitigs -> Align & Select -> Error Correct

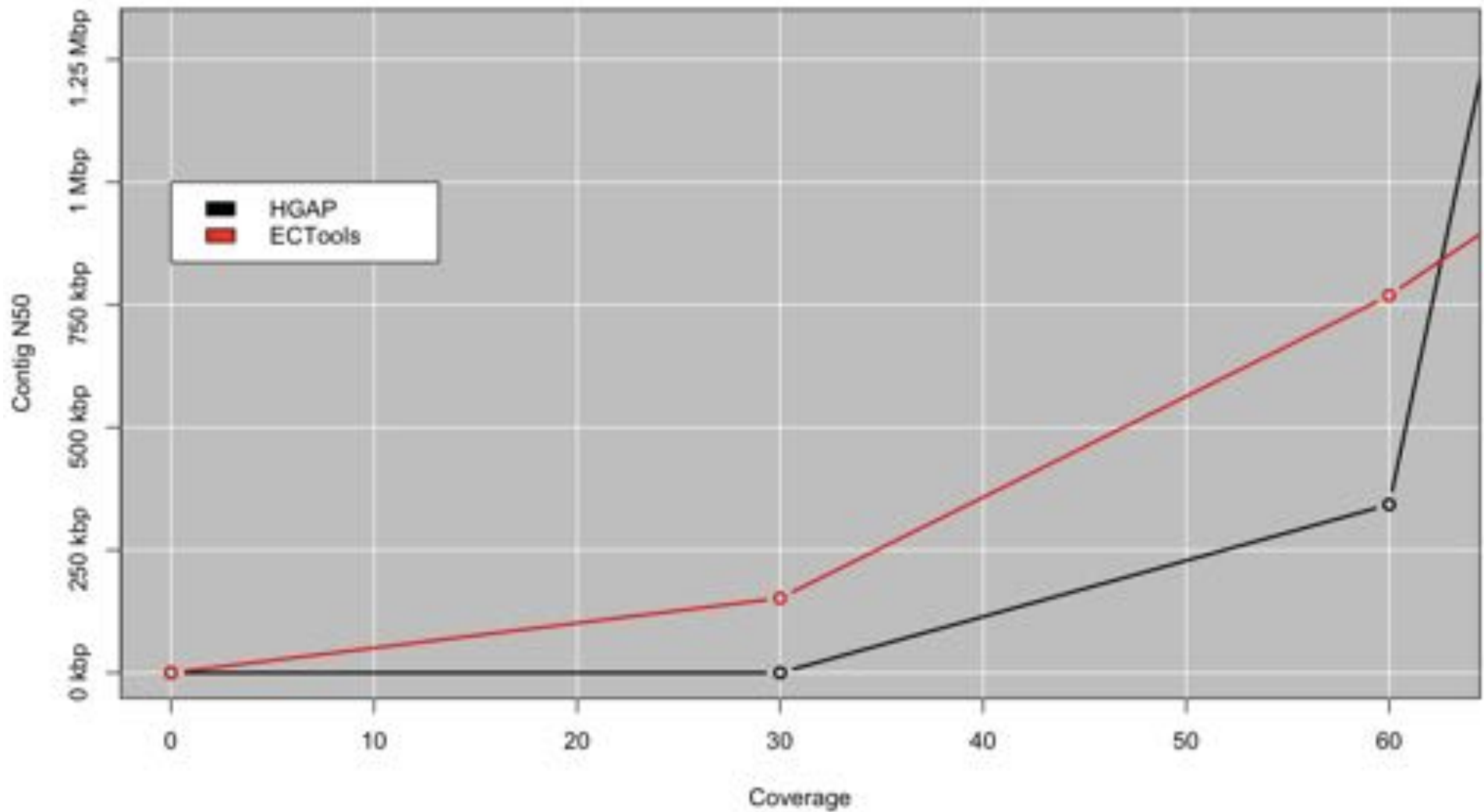
Can Help us overcome:

1. Error Dense Regions – Longer sequences have more seeds to match
2. Simple Repeats – Longer sequences easier to resolve

However, cannot overcome Illumina coverage gaps & other biases

A. thaliana Ler-0

<http://blog.pacificbiosciences.com/2013/08/new-data-release-arabidopsis-assembly.html>

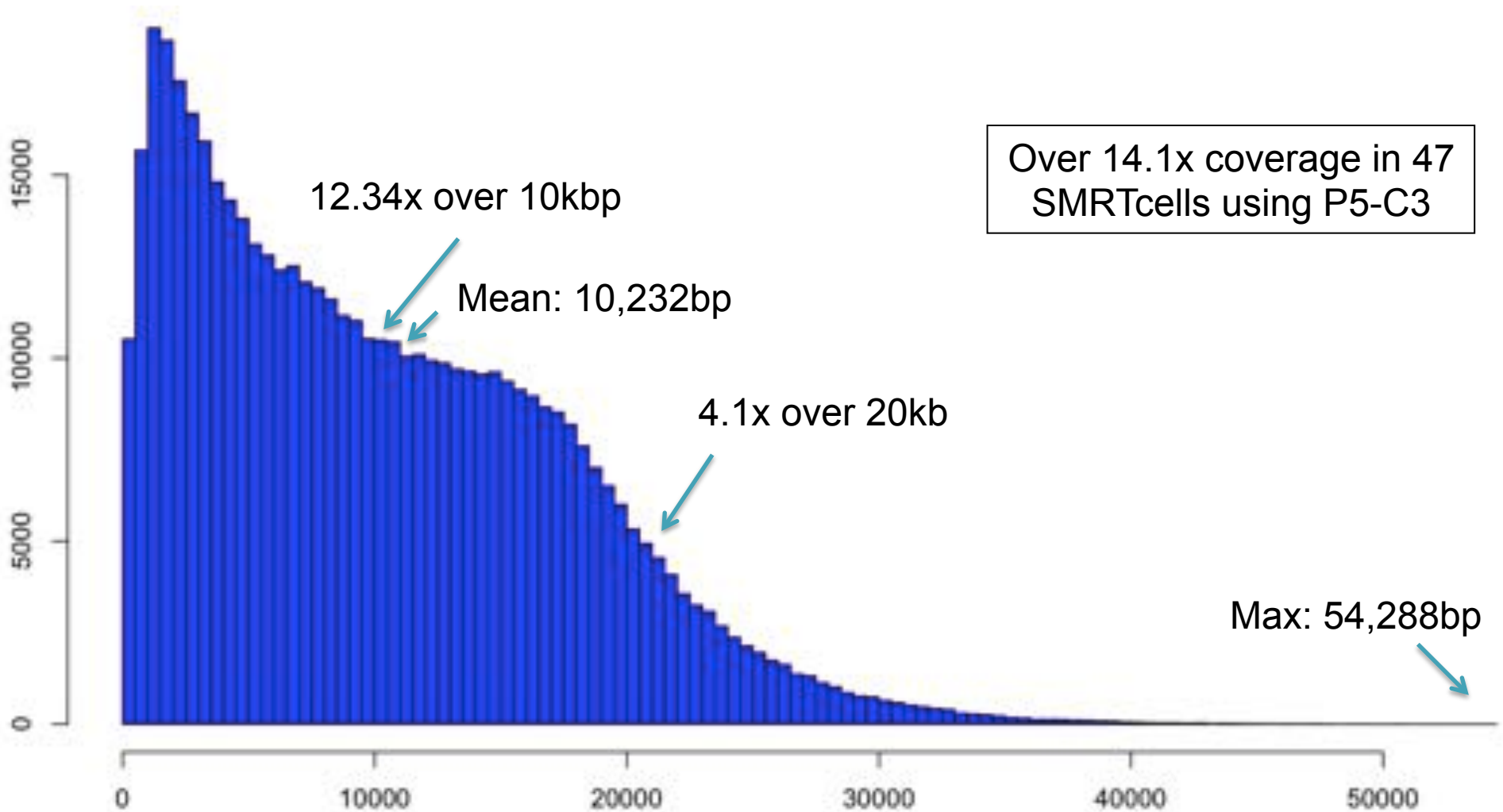


O. sativa pv Indica (IR64)



PacBio RS II sequencing at PacBio

- Size selection using an 10 Kb elution window on a BluePippin™ device from Sage Science

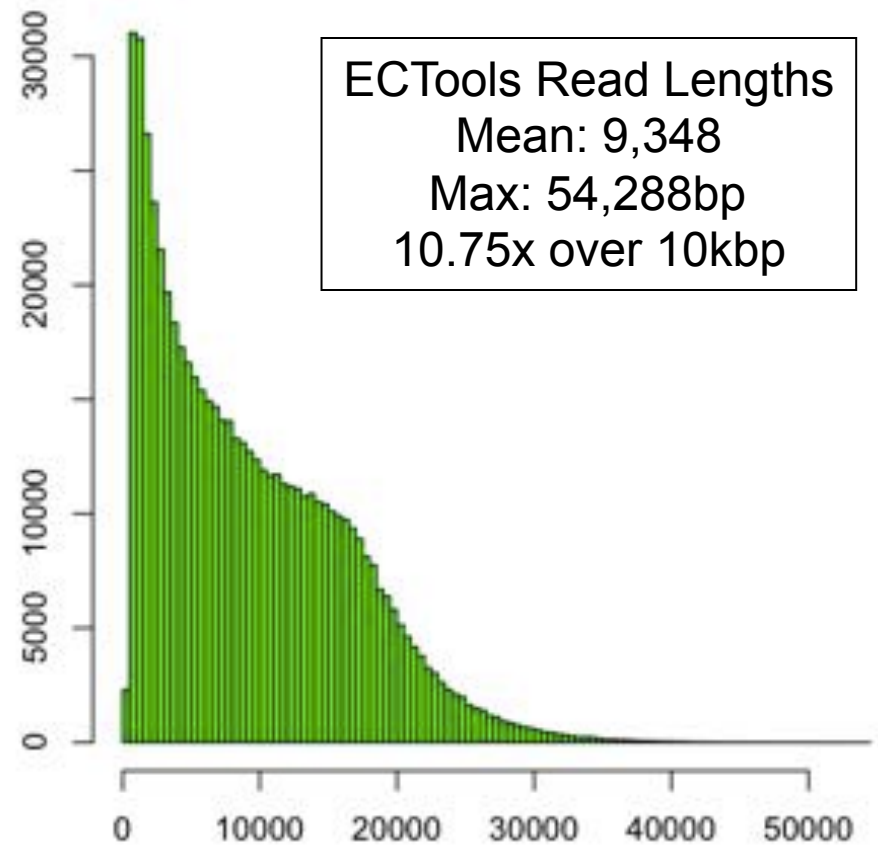


O. sativa pv Indica (IR64)

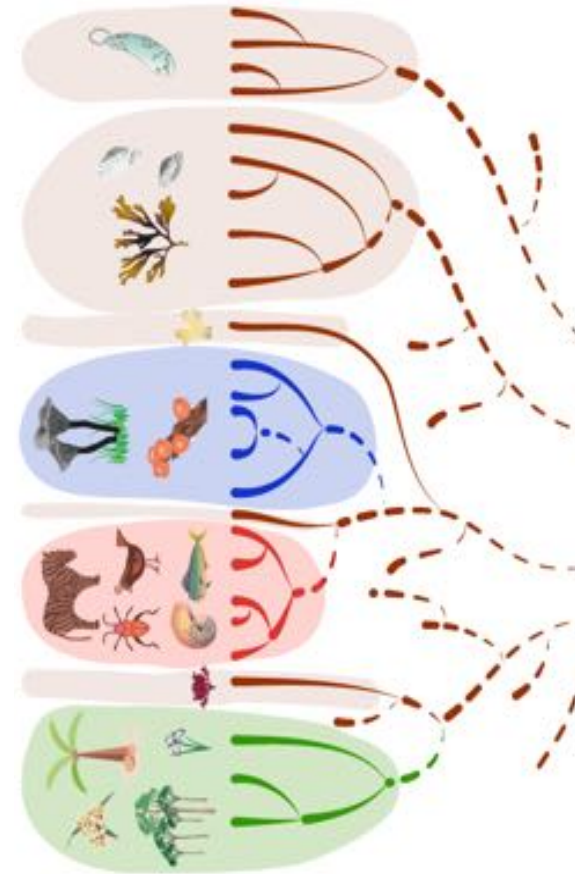
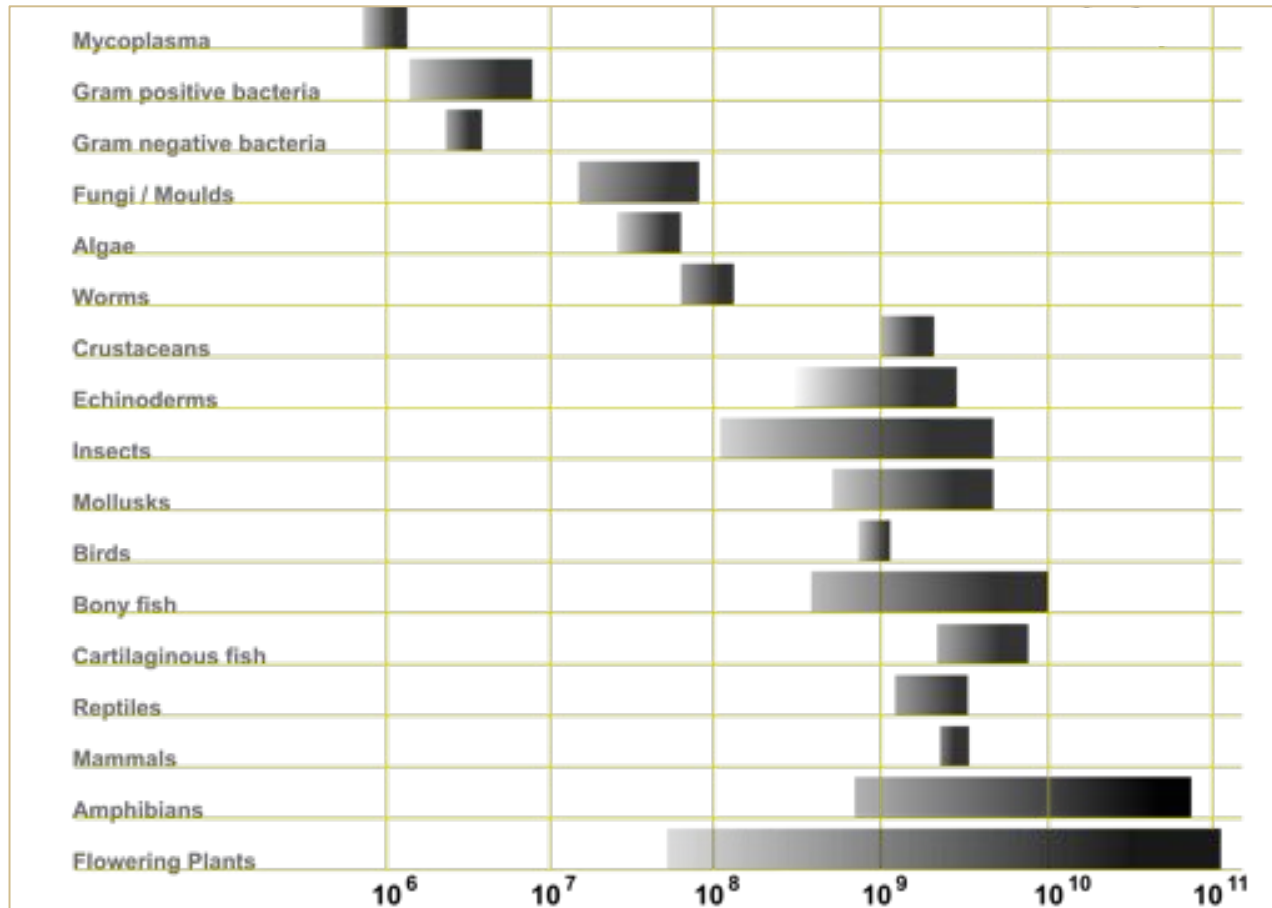
Genome size: ~370 Mb
Chromosome N50: ~29.7 Mbp



Assembly	Contig NG50
MiSeq Fragments 25x 456bp (3 runs 2x300 @ 450 FLASH)	19,078
“ALLPATHS-recipe” 50x 2x100bp @ 180 36x 2x50bp @ 2100 51x 2x50bp @ 4800	18,450
ECTools 10.7x @ 10kbp	271,885

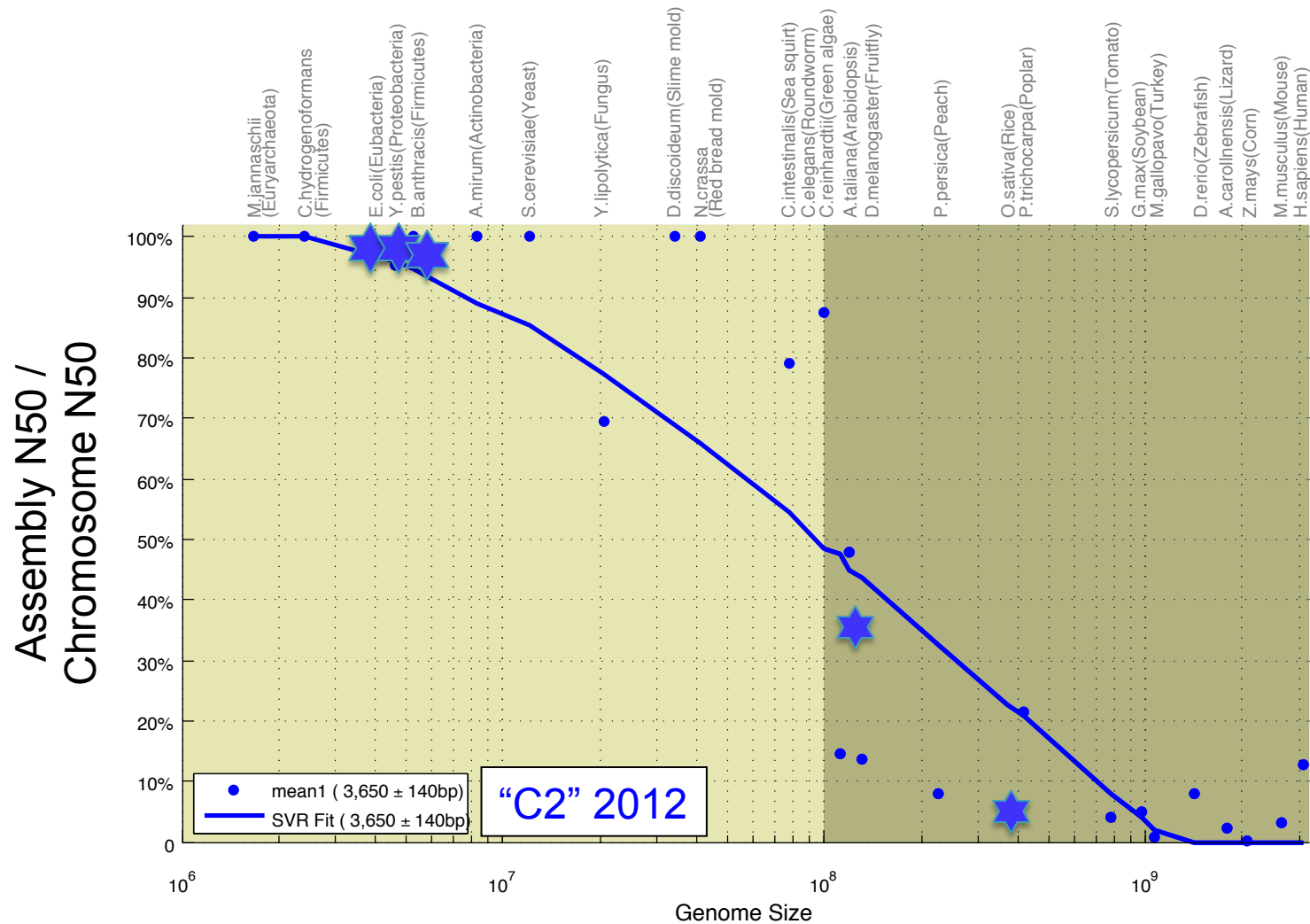


What should we expect from an assembly?



https://en.wikipedia.org/wiki/Genome_size

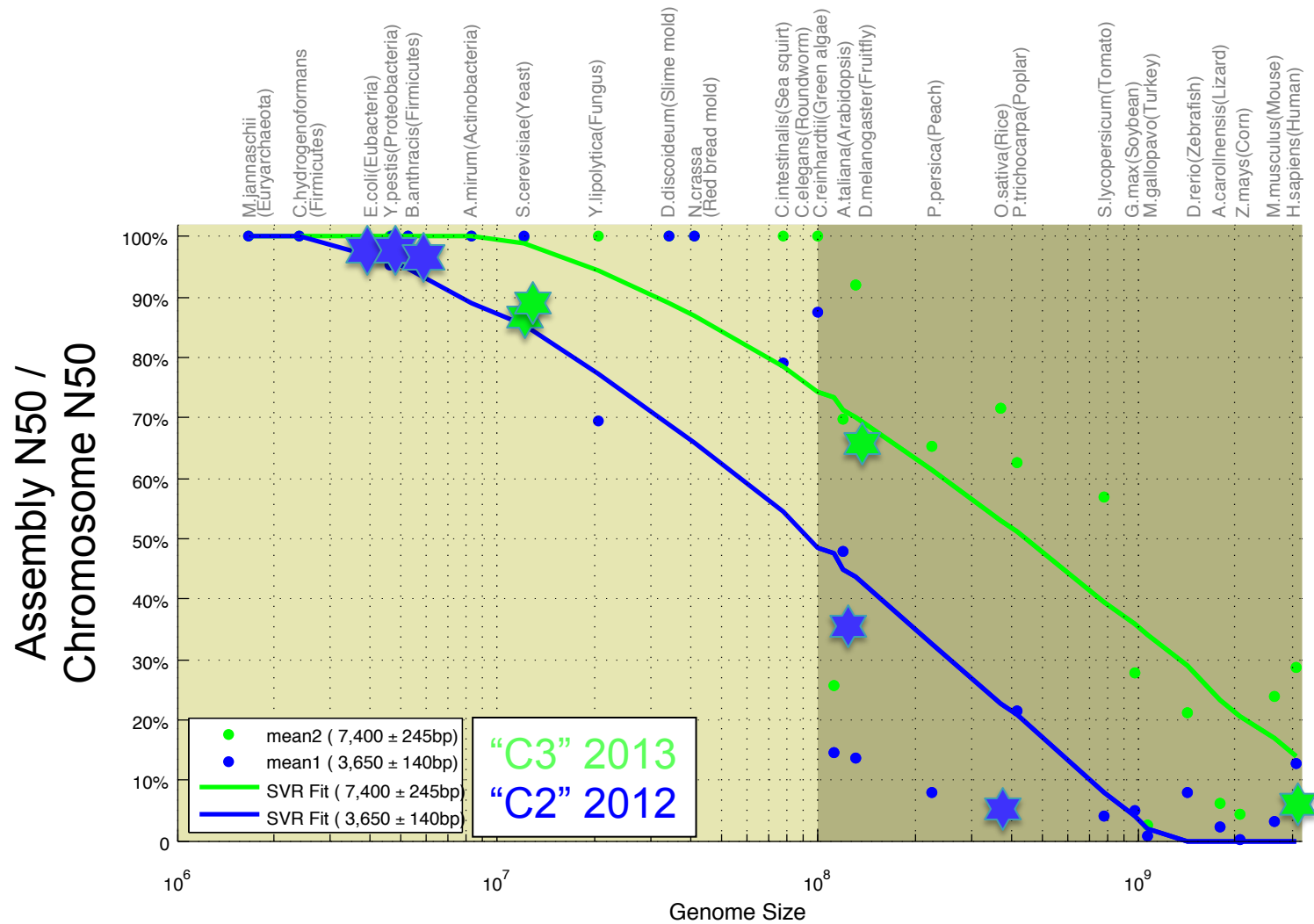
Assembly Complexity of Long Reads



Assembly complexity of long read sequencing

Lee, H*, Gurtowski, J*, Yoo, S, Marcus, S, McCombie, WR, Schatz MC (2014) *In preparation*

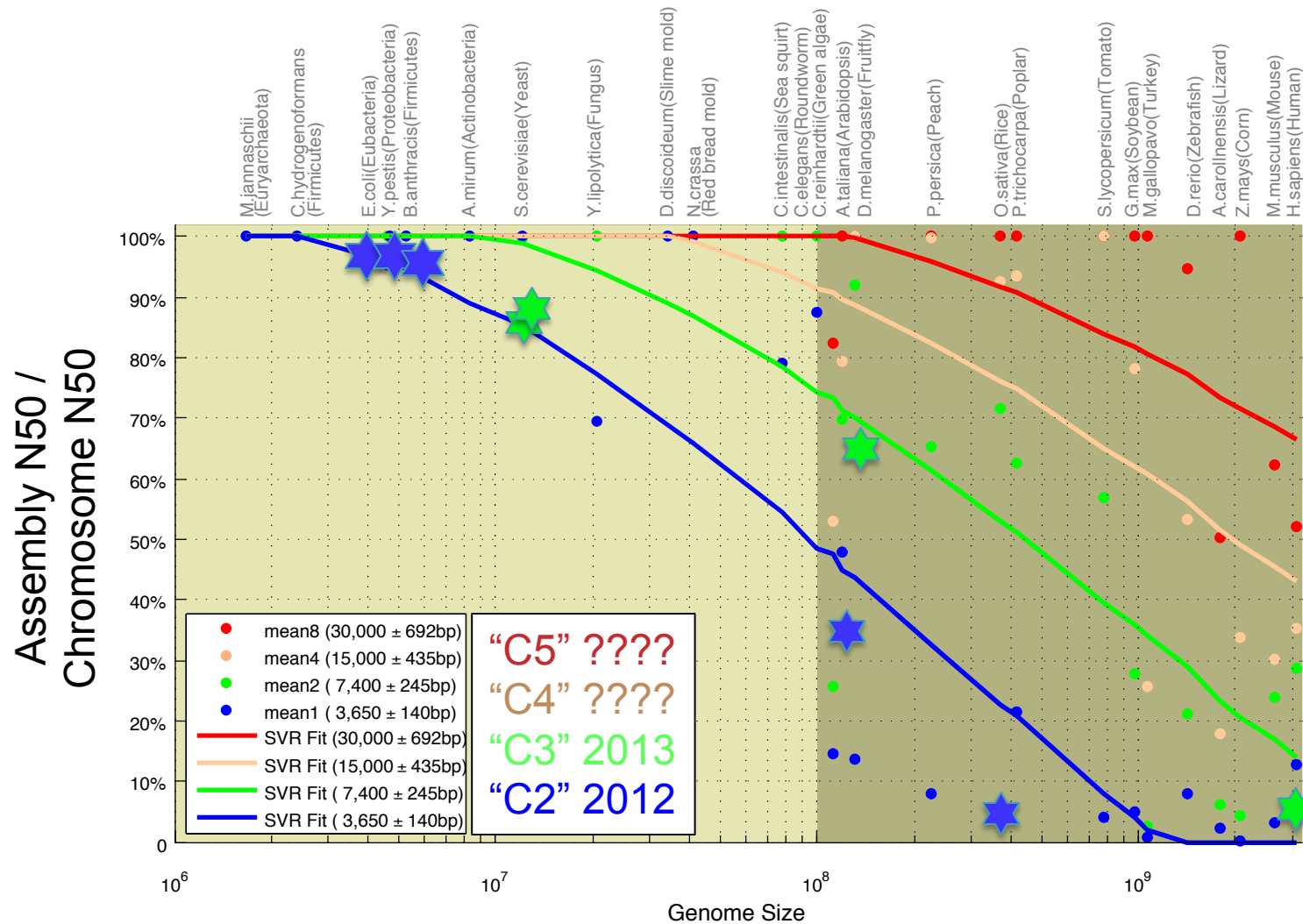
Assembly Complexity of Long Reads



Assembly complexity of long read sequencing

Lee, H*, Gurtowski, J*, Yoo, S, Marcus, S, McCombie, WR, Schatz MC (2014) *In preparation*

Assembly Complexity of Long Reads



Assembly complexity of long read sequencing

Lee, H*, Gurtowski, J*, Yoo, S, Marcus, S, McCombie, WR, Schatz MC. (2014) *In preparation*

Assembly Recommendations

- **Long read sequencing of eukaryotic genomes is here**

- **Recommendations**

- < 100 Mbp: HGAP/PacBio2CA @ 100x PB C3-P5
expect near perfect chromosome arms

- < 1GB: HGAP/PacBio2CA @ 100x PB C3-P5
expect high quality assembly: contig N50 over 1Mbp

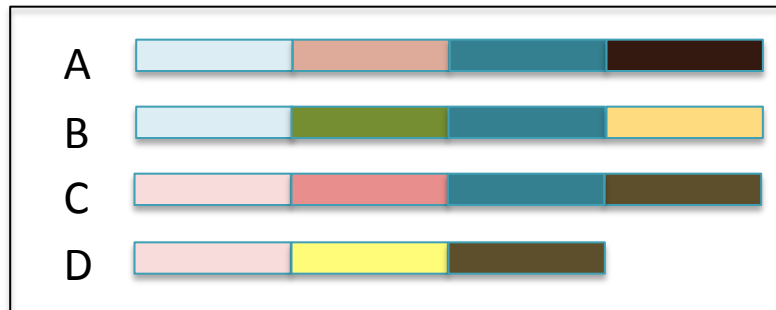
- > 1GB: hybrid/gap filling
expect contig N50 to be 100kbp – 1Mbp

- > 5GB: Email mschatz@cshl.edu

- **Caveats**

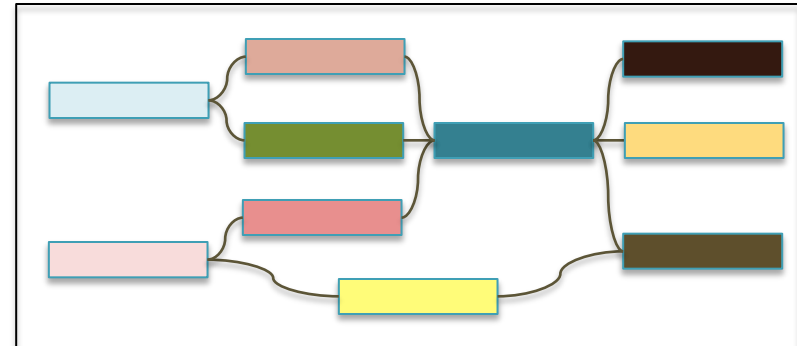
- Model only as good as the available references (esp. haploid sequences)
 - Technologies are quickly improving, exciting new scaffolding technologies

Pan-Genome Alignment & Assembly



Time to start considering problems for which N complete genomes is the input to study the “pan-genome”

- Available today for many microbial species, near future for higher eukaryotes



Pan-genome colored de Bruijn graph

- Encodes all the sequence relationships between the genomes
- How well conserved is a given sequence?
- What are the pan-genome network properties?

Rapid pan genome analysis with suffix skips

Marcus, S, Schatz, MC (2014) *In preparation*

Outline

1. De novo assembly by analogy
2. Long Read Assembly
3. **Disease Analytics**



Scalpel: Haplotype Microassembly

DNA sequence **micro-assembly** pipeline for accurate detection and validation of *de novo* mutations (SNPs, indels) within exome-capture data.



Features

1. Combine **mapping** and **assembly**
2. Exhaustive search of **haplotypes**
3. **De novo mutations**



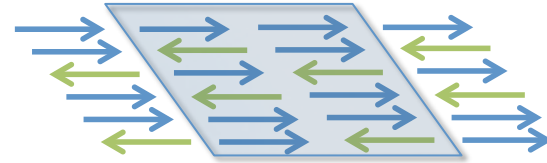
NRXN1 *de novo* SNP
(auSSC12501 chr2:50724605)

Accurate detection of de novo and transmitted INDELs within exome-capture data using micro-assembly

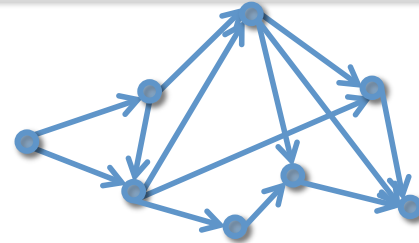
Narzisi, G, O’Rawe, J, Iossifov, I, Lee, Y, Wang, Z, Wu, Y, Lyon, G, Wigler, M, Schatz, MC (2014) *Under review.*

Scalpel Pipeline

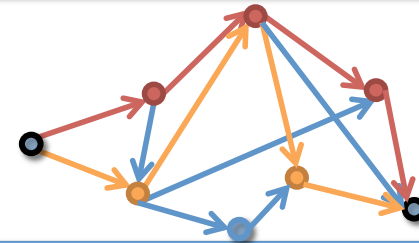
Extract reads mapping within the exon including (1) well-mapped reads, (2) soft-clipped reads, and (3) anchored pairs



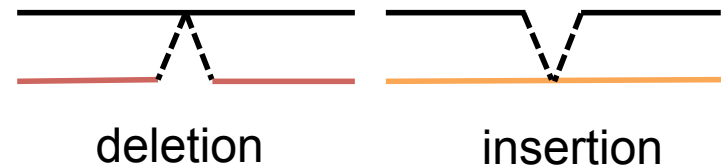
Decompose reads into overlapping k -mers and construct de Bruijn graph from the reads



Find end-to-end haplotype paths spanning the region

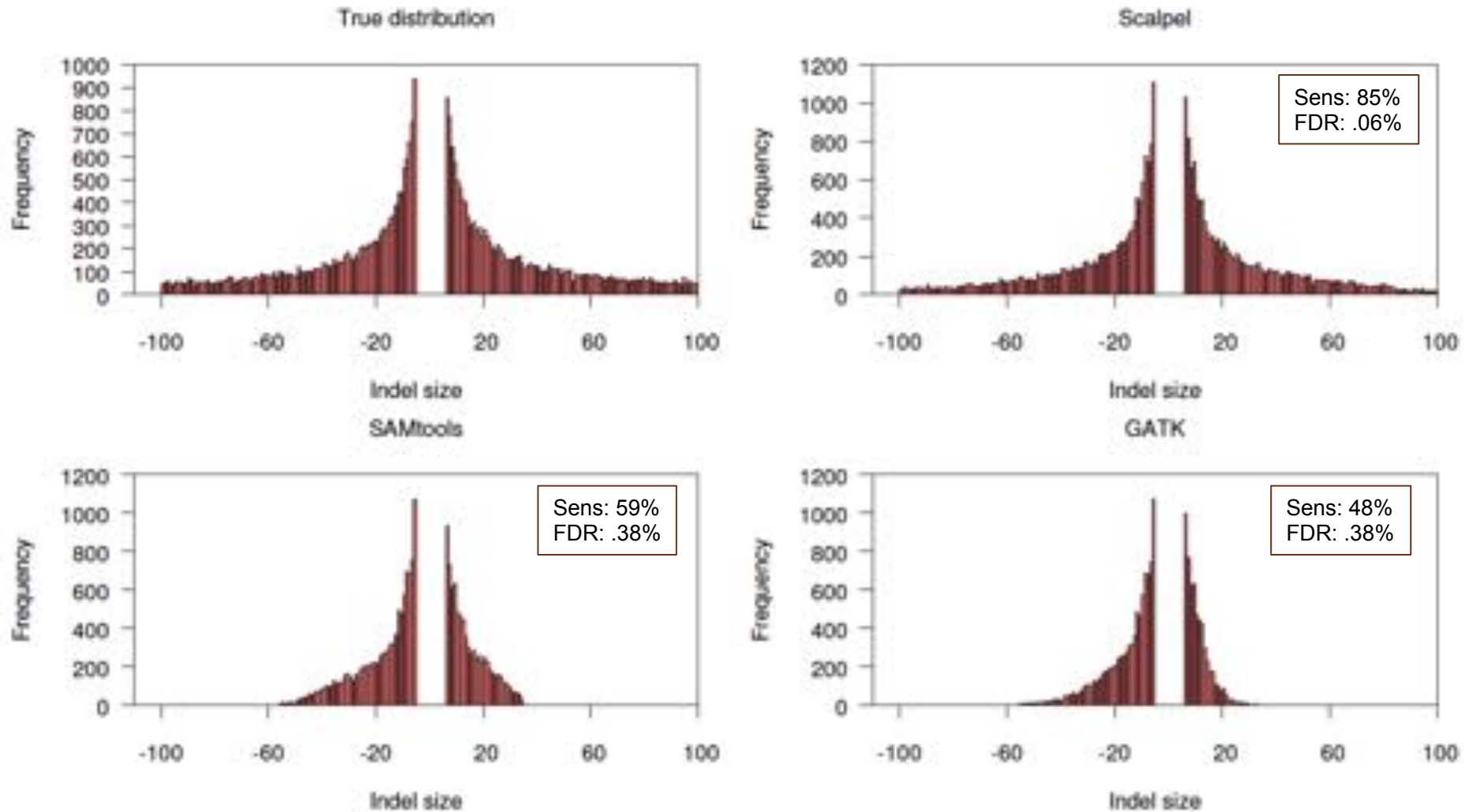


Align assembled sequences to reference to detect mutations



Simulation Analysis

Indel size distribution (length > 5 bp)



Simulated 10,000 indels in an exome from a known log-normal distribution

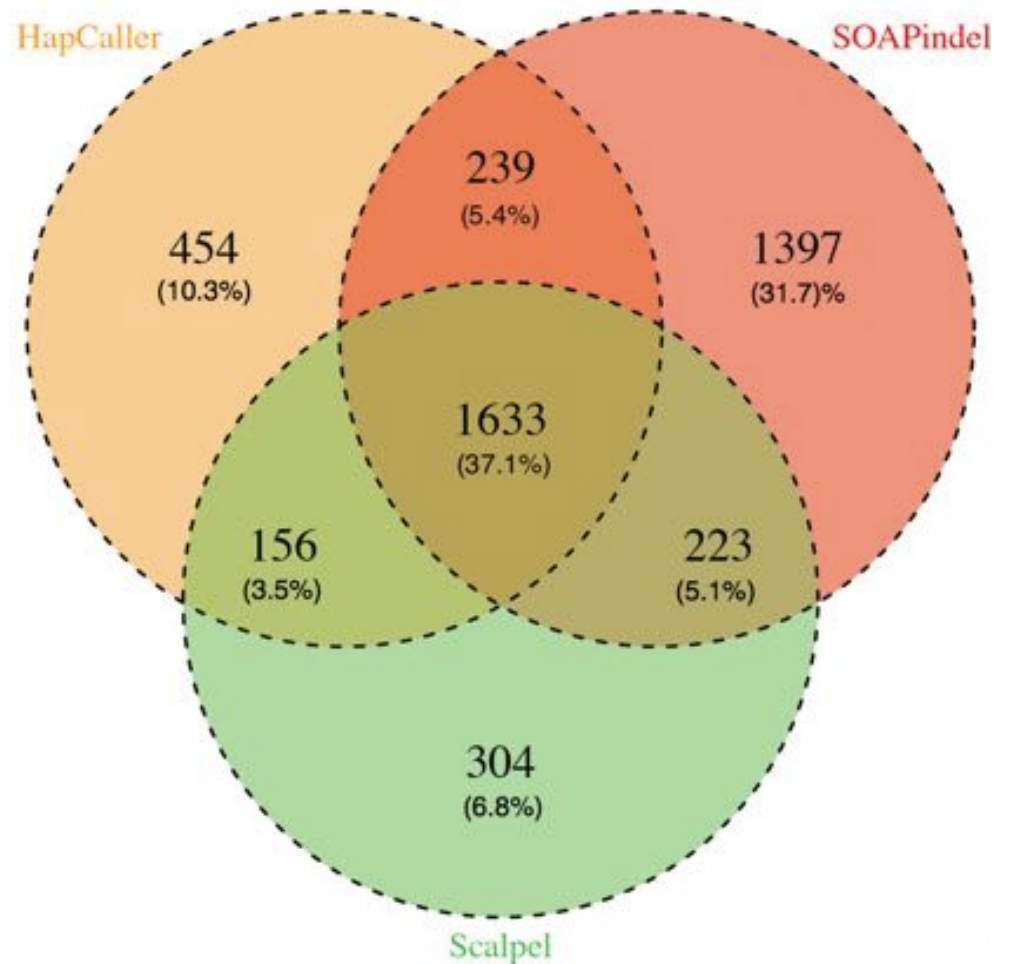
Experimental Analysis & Validation

Selected one deep coverage exome for deep analysis

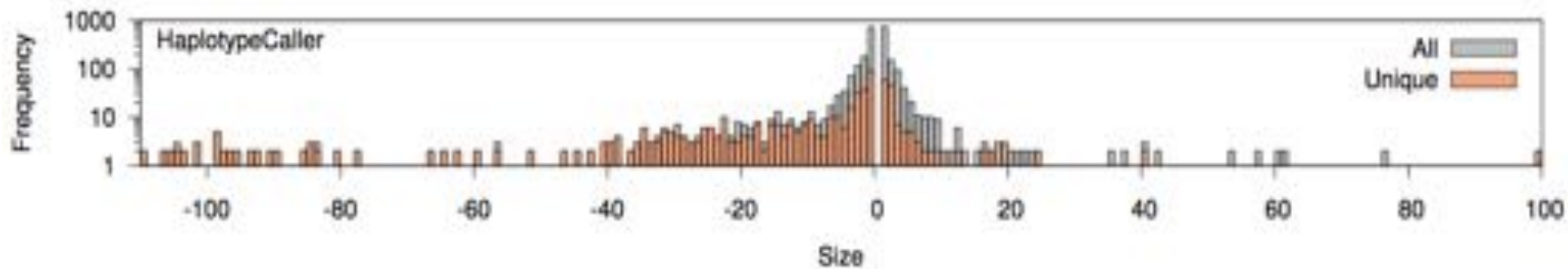
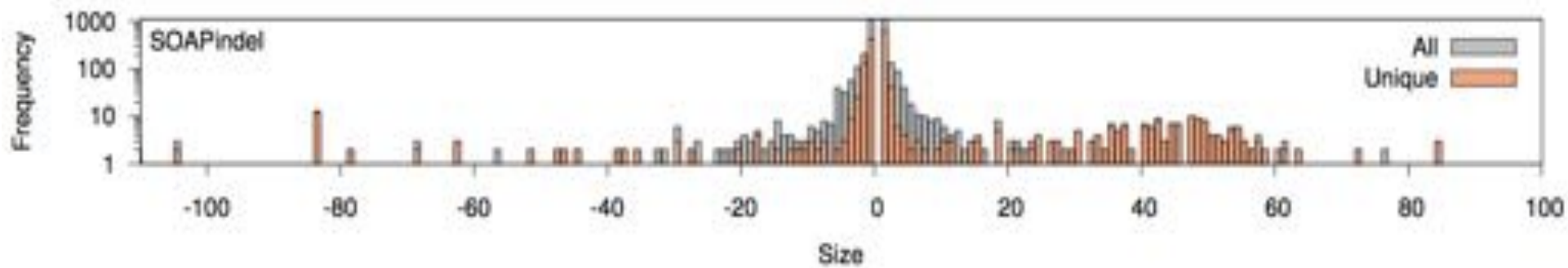
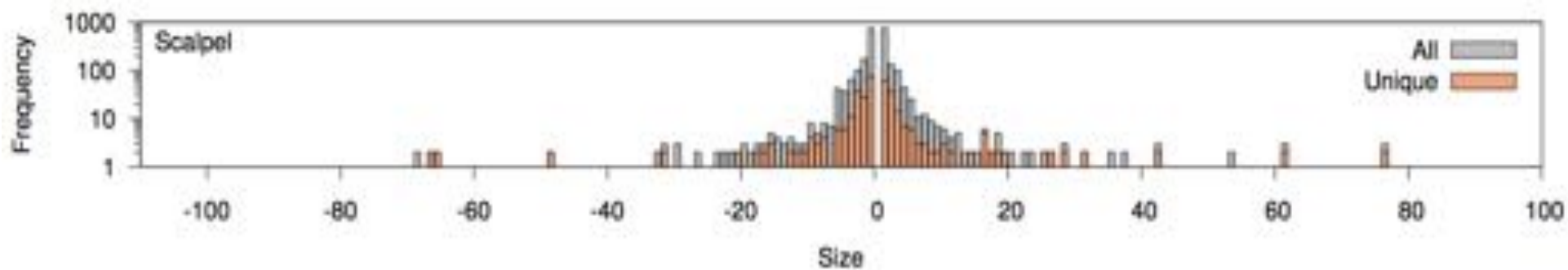
- Individual was diagnosed with ADHD and turrets syndrome
- 80% of the target at >20x coverage
- Evaluated with Scalpel, SOAPindel, and GATK Haplotype Caller

1000 indels selected for validation

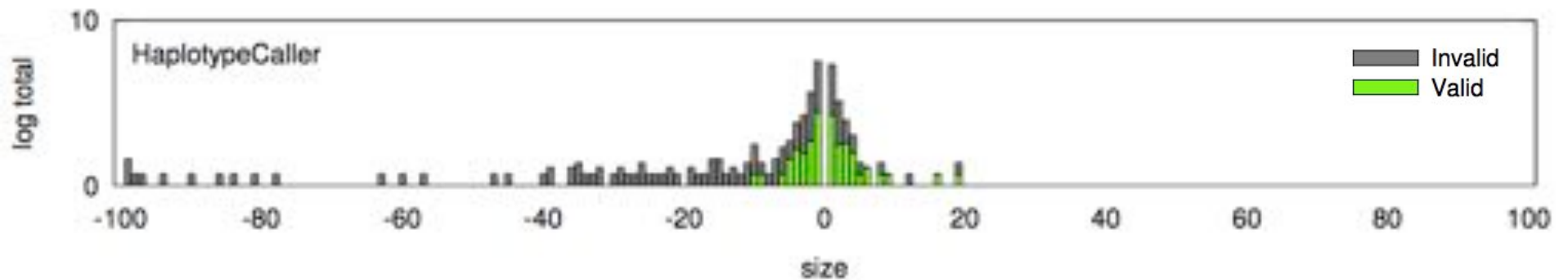
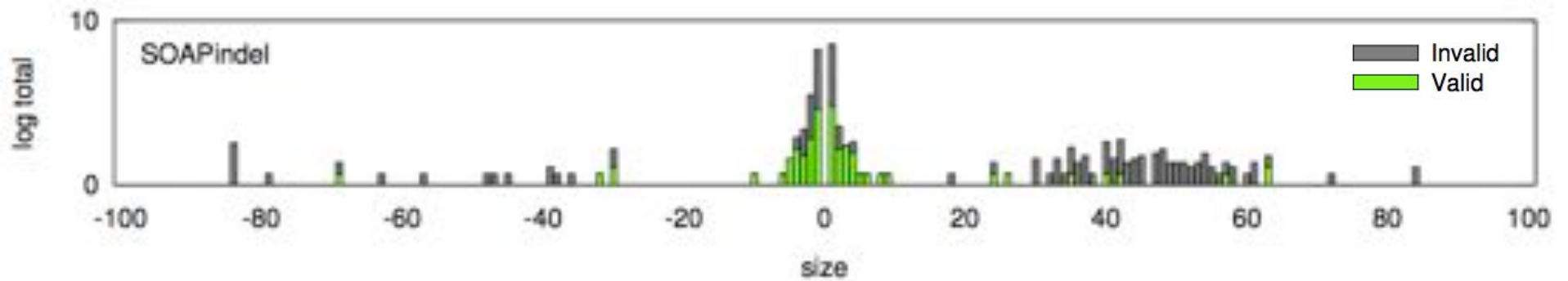
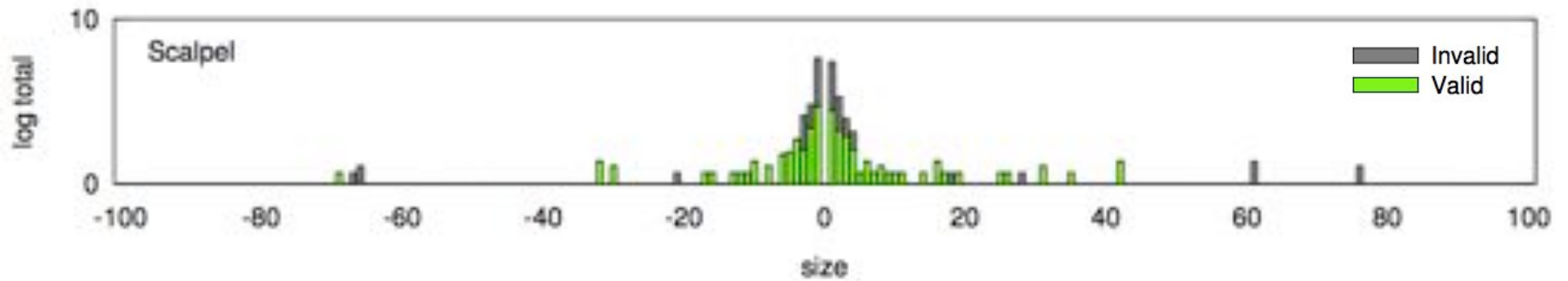
- 200 Scalpel
- 200 GATK Haplotype Caller
- 200 SOAPindel
- 200 within the intersection
- 200 long indels (>30bp)



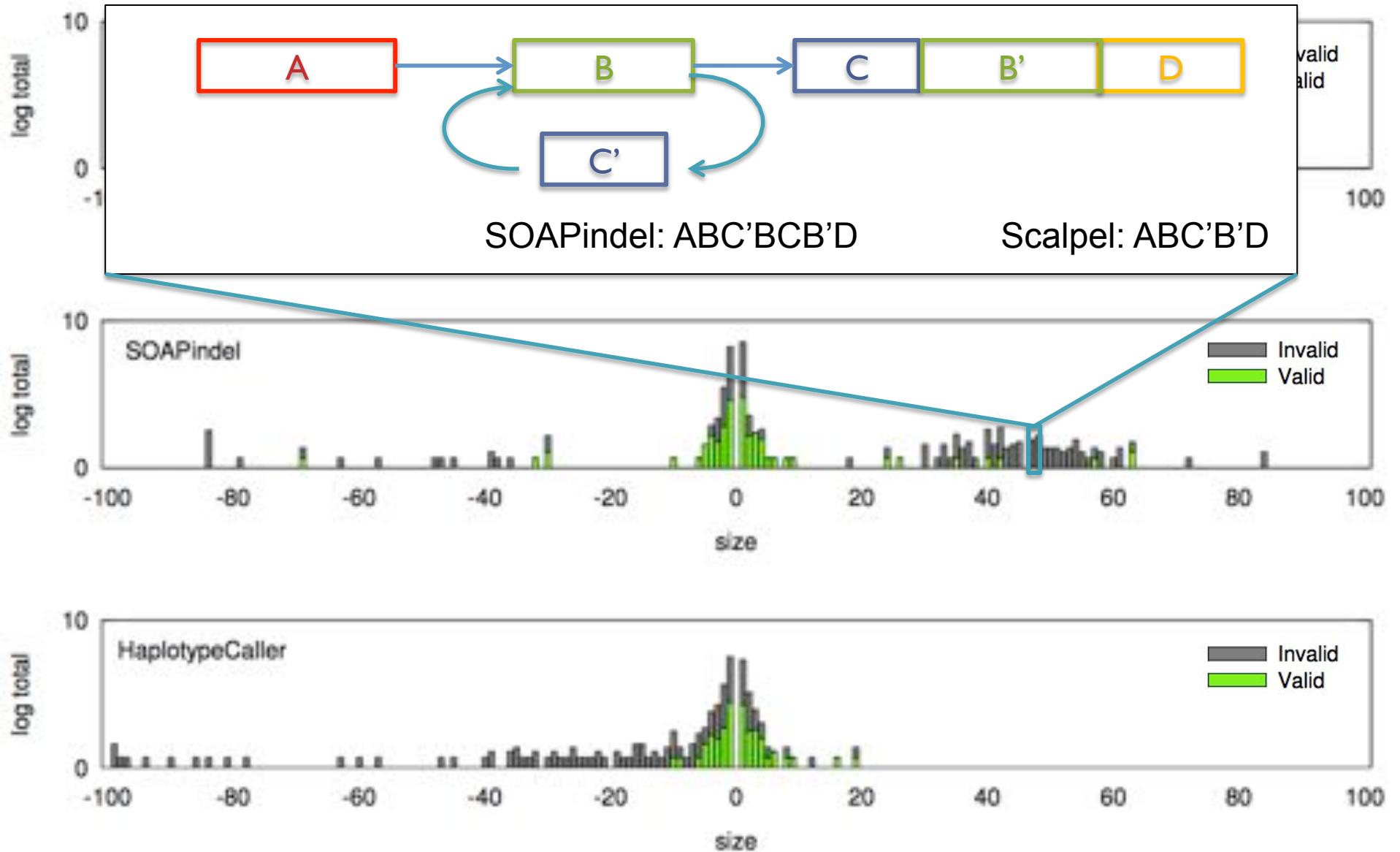
Scalpel Indel Discovery



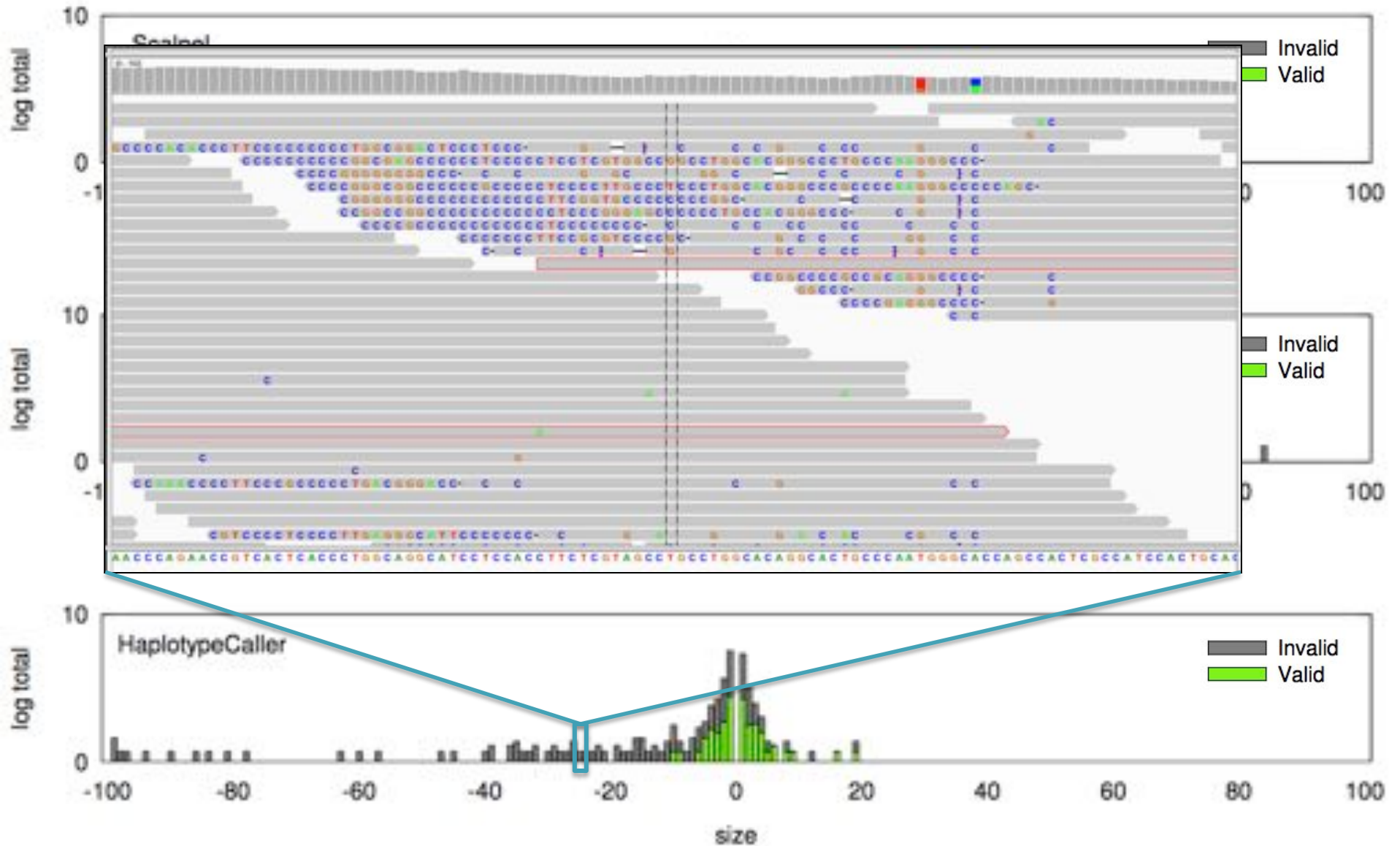
Scalpel Indel Discovery



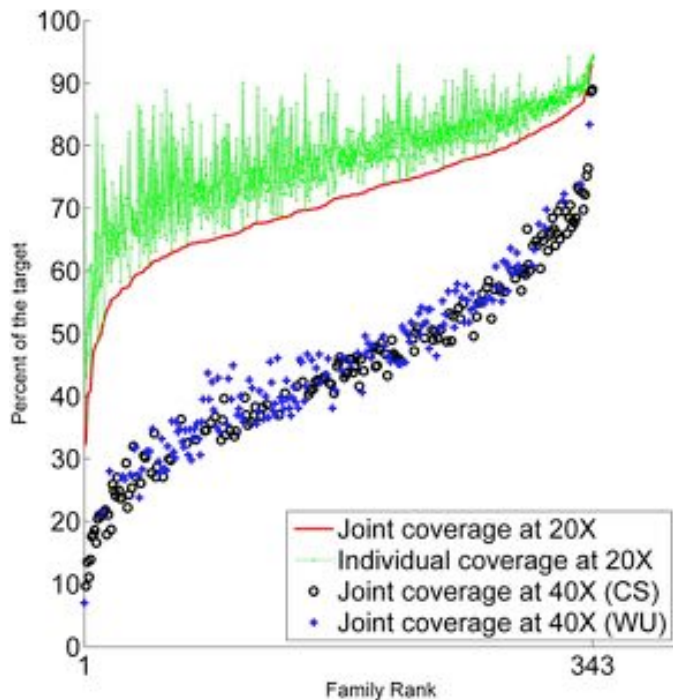
Scalpel Indel Discovery



Scalpel Indel Discovery



Exome sequencing of the SSC



Last year saw 3 reports of >593 families from the Simons Simplex Collection

- Parents plus one child with autism and one non-autistic sibling
- All attempted to find mutations enriched in the autistic children
- Iossifov (343) and O’Roak (50) used GATK, Sanders (200) didn’t attempt to identify indels

De novo gene disruptions in children on the autism spectrum

Iossifov *et al.* (2012) *Neuron*. 74:2 285-299

De novo mutations revealed by whole-exome sequencing are strongly associated with autism

Sanders *et al.* (2012) *Nature*. 485, 237–241.

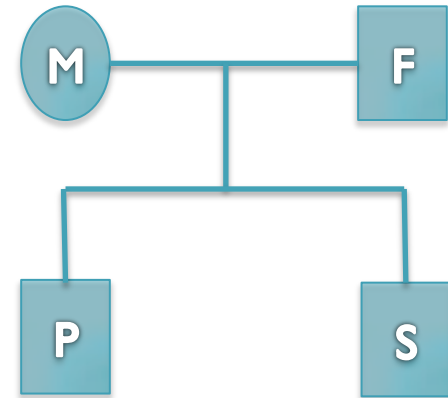
Sporadic autism exomes reveal a highly interconnected protein network of de novo mutations

O’Roak *et al.* (2012) *Nature*. 485, 246–250.

De novo mutation discovery and validation

Concept: Identify mutations not present in parents.

Challenge: Sequencing errors in the child or low coverage in parents lead to false positive de novos



Reference: . . . TCAAATCCTTTTAAATAAGAAGAGCTGACA . . .

Father: . . . TCAAATCCTTTTAAATAAGAAGAGCTGACA . . .

Mother: . . . TCAAATCCTTTTAAATAAGAAGAGCTGACA . . .

Sibling: . . . TCAAATCCTTTTAAATAAGAAGAGCTGACA . . .

Proband(1): . . . TCAAATCCTTTTAAATAAGAAGAGCTGACA . . .

Proband(2): . . . TCAAATCCTTTTAAAT****AAGAGCTGACA . . .

4bp heterozygous deletion at chr15:9352406 | CHD2

De novo Genetics of Autism

- In 593 family quads so far, we see significant enrichment in de novo **likely gene killers** in the autistic kids
 - Overall rate basically 1:1
 - 2:1 enrichment in nonsense mutations
 - 2:1 enrichment in frameshift indels
 - 4:1 enrichment in splice-site mutations
 - Most de novo originate in the paternal line in an age-dependent manner (56:18 of the mutations that we could determine)
- Observe strong overlap with fragile X protein (FMR1) network
 - Related to neuron development and synaptic plasticity
 - Also strong overlap with chromatin remodelers

Accurate detection of de novo and transmitted INDELs within exome-capture data using micro-assembly

Narzisi, G, O’Rawe, J, Iossifov, I, Lee, Y, Wang, Z, Wu, Y, Lyon, G, Wigler, M, Schatz, MC (2014) *Under review.*

Summary

Biotechnology

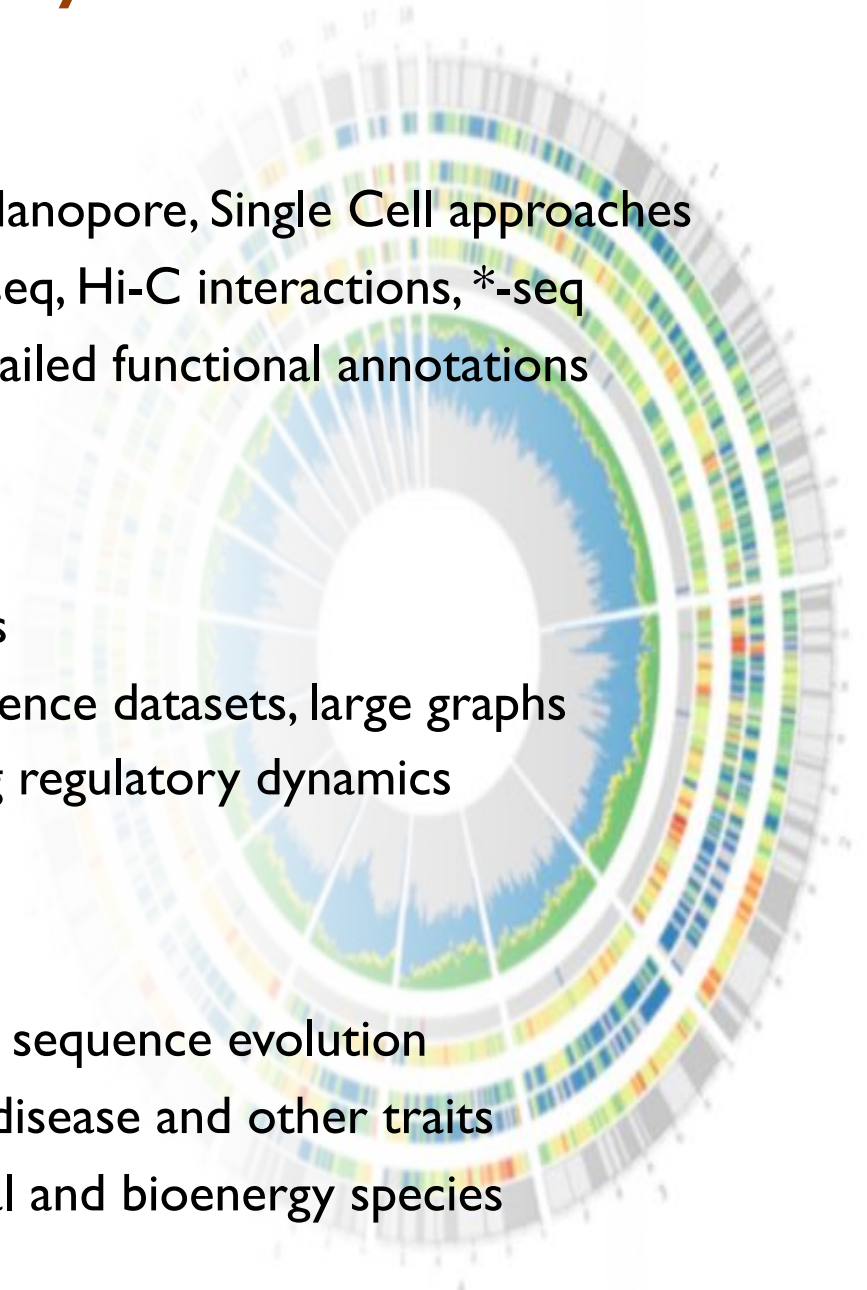
- Sequencing: Illumina, PacBio, Oxford Nanopore, Single Cell approaches
- Biochemical assays: RNA-seq, Methyl-seq, Hi-C interactions, *-seq
- More accurate assemblies & more detailed functional annotations

Algorithmics

- Highly scalable algorithms and systems
- Indexing and analyzing very large sequence datasets, large graphs
- Constructing Pan-genomes & inferring regulatory dynamics

Comparative Genomics

- Cross species comparisons, models of sequence evolution
- Identifying mutations associated with disease and other traits
- Genotype-to-phenotype of agricultural and bioenergy species



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Wigler Lab

Lyon Lab

Hannon Lab

Gingeras Lab

Jackson Lab

Hicks Lab

Iossifov Lab

Levy Lab

Lippman Lab

Martienssen Lab

Tuveson Lab

Ware Lab

Pacific Biosciences

SFARI

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Institute



U.S. DEPARTMENT OF
ENERGY



**Biological Data Sciences
Cold Spring Harbor Laboratory, Nov 5 - 8, 2014**



Thank you

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